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Genetic Determinants of Cortical Structure (Thickness, Surface Area and Volumes) among Disease Free Adults in the CHARGE Consortium

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Abstract

Cortical thickness, surface area and volumes (MRI cortical measures) vary with age and cognitive function, and in neurological and psychiatric diseases. We examined heritability, genetic correlations and genome-wide associations of cortical measures across the whole cortex, and in 34 anatomically predefined regions. Our discovery sample comprised 22,824 individuals from 20 cohorts within the Cohorts for Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium and the United Kingdom Biobank. Significant associations were replicated in the Enhancing Neuroimaging Genetics through Meta-analysis (ENIGMA) consortium, and their biological implications explored using bioinformatic annotation and pathway analyses. We identified genetic heterogeneity between cortical measures and brain regions, and 160 genome-wide significant associations pointing to wnt/β-catenin, TGF-β and sonic hedgehog pathways. There was enrichment for genes involved in anthropometric traits, hindbrain development, vascular and neurodegenerative disease and psychiatric conditions. These data are a rich resource for studies of the biological mechanisms behind cortical development and aging.

Introduction

The cortex is the largest part of the human brain, associated with higher brain functions such as perception, thought and action. Brain cortical thickness (CTh), cortical surface area (CSA) and cortical volume (CV) are morphological markers of cortical structure obtained from magnetic resonance imaging (MRI). These measures change with age¹⁻³ and are linked to cognitive functioning^{4,5}. The human cortex is also vulnerable to a wide range of disease or pathologies, ranging from developmental disorders and early onset psychiatric and neurological diseases to neurodegenerative conditions manifesting late in life. Abnormalities in global or regional CTh, CSA and CV have been observed in neurological and psychiatric disorders such as Alzheimer's disease^{6,7}, Parkinson's disease^{8,9}, multiple sclerosis^{10,11}, schizophrenia^{12,13}, bipolar disorder^{12,14,15}, depression^{15,16} and autism^{17,18}. The best method to study human cortical structure during life is using brain MRI. Hence, understanding the genetic determinants of the most robust MRI cortical markers in apparently normal adults could identify biological pathways relevant to brain development, aging and various diseases. Neurons in the neocortex are organized in columns which run perpendicular to the surface of the cerebral cortex¹⁹; and, according to the radial unit hypothesis. CTh is determined by the number of cells within the columns and CSA is determined by the number of columns²⁰. Thus, CTh and CSA reflect different mechanisms in cortical development²⁰⁻²⁴ and are likely influenced by different genetic factors^{25,26}. CV, which is the product of CTh and CSA, is determined by a combination of these two measures, but the relative contribution of CTh and CSA to CV may vary across brain regions. CTh, CSA and CV are all strongly heritable traits^{22,24-30} with estimated heritability of 0.69 to 0.81 for global CTh, and from 0.42 to 0.90 for global CSA²⁴⁻²⁶. Across different cortical regions however, there is substantial regional variation in heritability of CTh, CSA and CV^{22,24-30}. Since CTh, CSA and CV are differentially heritable and genetically heterogeneous, we explored the genetics of each of

these imaging markers using genome-wide association analyses in large population-based samples (GWAS). We studied CTh, CSA and CV in the whole cortex and in 34 cortical regions in 22,824 individuals from 21 discovery cohorts and replicated the strongest associations in 22,363 persons from the ENIGMA consortium.

Results

Genome-wide association analysis

Global Cortical Measures: The analyses of global CTh, CSA and CV included 22,163, 18,617 and 22,824 individuals respectively. After a conservative correction for multiple testing (p_{discovery}<1.09x10⁻⁹), we identified no significant associations with global CTh. However, we identified 12 independent loci associated with global CSA (n=6) and CV (n=6). These are displayed in Table S8 and Supplementary Figures 1 and 2. Five of the 6 CSA loci were replicated in an external (ENIGMA consortium) sample³¹. The ENIGMA consortium only analyzed CSA and CTh.

Regional Cortical Measures: GWAS of CTh, CSA and CV in 34 cortical regions of interest (ROIs) identified 148 significant associations. There were 16 independent loci across 8 chromosomes determining CTh of 9 regions (Table S9), 54 loci across 16 chromosomes associated with CSA of 21 regions (Table S10), and 78 loci across 17 chromosomes determining CV of 23 cortical regions (Table S11). We replicated 57 out of 70 regional CTh and CSA loci in the ENIGMA consortium sample³¹ using a conservative replication threshold of p_{Replication}=3.1x10⁻⁴, 0.05/160. Region-specific variants with the strongest association at each genomic locus are shown in Tables 1-3. Chromosomal ideograms showing genomewide significant associations with global and regional cortical measures in the discovery stage are presented in Figure 1.

The strongest associations with CTh and CV were observed for rs2033939 at 15q14 (p_{Discovery}, CTh=1.17x10⁻⁷³ and p_{Discovery}, CV=4.34x10⁻¹³³) in the postcentral (primary somatosensory) cortex, and for CSA with rs1080066 at 15q14 (p_{Discovery}, CSA=8.45x10⁻¹⁰⁹) in the precentral (primary motor) cortex. Figure 2 shows the lowest p-value of each cortical region. The postcentral cortex was also the region with the largest number of independent associations, mainly at a locus on 15q14. The corresponding regional association plots are presented in

Supplementary Figure 3.

QQ plots of all meta-analyses are presented in Supplementary Figures 4-7.

Associations across Cortical Measures and with Other Traits: Table S12 presents variants which are associated with the CSA or the CV across multiple regions. We observed 25 SNPs that determined both the CSA and CV of a given region, 4 SNPs that determined CTh and CV of the same region, but no SNPs that determined both the CTh and CSA of any given region (Table S13). In the UK Biobank³², 3 SNPs were associated with the same cortical measure at a given cortical region as in our study. These included associations between rs2033939 at 15q14 and postcentral CTh, rs2279829 at 3q24 and CSA of the pars triangularis and postcentral gyrus and rs76341705 at 14q23.1 and CSA of lingual gyrus, respectively (Table S14). When assessing genetic overlap with other traits, we observed that SNPs determining these cortical measures have been previously associated with anthropometric (height), neurologic (Parkinson's disease, corticobasal degeneration, Alzheimer's disease), psychiatric (neuroticism, schizophrenia) and cognitive performance traits as well as with total intracranial volume (TIV) on brain MRI (Tables S15-S17).

Gene Identification

Positional mapping based on ANNOVAR showed that most of the lead SNPs were intergenic and intronic (Figure 3). One variant, rs2279829, which was associated with both CSA and CV of the pars triangularis, postcentral and supramarginal cortices, is located in the 3'prime UTR of *ZIC4* at 3q24. We also found an exonic variant, rs10283100, in gene *ENPP2* at 8q24.12 associated with CV of the insula.

We used multiple strategies beyond positional annotation to identify specific genes implicated by the various GWAS associated SNPs. FUMA identified 232 genes whose

expression was determined by these variants (eQTL) and these and other genes implicated by chromatin interaction mapping are shown in Tables S18–S20. MAGMA gene-based association analyses revealed 70 significantly associated (p<5.87*10-8) genes (Tables S21–S23). For global CSA and CV, 7 of 9 genes associated with each measure overlapped, but there was no overlap with global CTh. For regional CSA and CV we found 28 genes across 13 cortical regions that determined both measures in the same region. Figure 4 summarizes the results of GTEx eQTL, chromatin interaction, positional annotation and gene-based mapping strategies for all regions. While there are overlapping genes identified using different approaches, only *DAAM1* gene (Chr14q23.1) is identified by all types of gene mapping for CV of insula. eQTL associations of our independent lead SNPs in the Religious Orders Study- Memory and Aging Project dorsolateral frontal cortex gene expression dataset are presented in Table S24.

Pathway analysis

MAGMA gene set analyses identified 7 pathways for CTh, 3 pathways for CSA and 9 pathways for CV (Table S25). Among them are the Gene Ontology (GO) gene sets 'hindbrain morphogenesis' (strongest association with thickness of middle temporal cortex), 'forebrain generation of neurons' (with surface area of precentral cortex), and 'central nervous system neuron development' (with volume of transverse temporal cortex). However, after Bonferroni correction only one significant pathway (p<1.02×10⁻⁷) remained: 'regulation of catabolic process' for CTh of inferior temporal cortex. InnateDB pathway analyses of genes mapped to independent lead SNPs by FUMA showed a significant overlap between CTh and CSA genes and the Wnt signaling pathway (Supplementary Figures 8 and 9) as well as a significant overlap between CV genes and the basal cell carcinoma pathway (Supplementary Figure 10).

Heritability

Heritability estimates (h^2) of global CTh were 0.64 (se=0.12; p=3×10⁻⁷) in ASPS-Fam and 0.45 (se=0.08; p=2.5x10⁻⁷) in RS. For CSA, h^2 was 0.84 (se=0.12; p=2.63×10⁻¹¹) in ASPS-Fam and 0.33 (se=0.08, p=1×10⁻⁴) in RS, and for CV, h^2 was 0.80 (se=0.11; p=1.10×10⁻⁹) in ASPS-Fam and 0.32 (se=0.08; p=1x10⁻⁴) in RS. There was a large range in heritability estimates of regional CTh, CSA and CV (Table S26).

Heritability based on common SNPs as estimated with LDSR was 0.25 (se=0.03) for global CTh, 0.29 (se=0.04) for global CSA and 0.30 (se=0.03) for global CV. LDSR heritability estimates of regional CTh, CSA and CV are presented in Table S26 and Supplementary Figure 11. For the regional analyses, the estimated heritability ranged from 0.05 to 0.18 for CTh, from 0.07 to 0.36 for CSA and from 0.06 to 0.32 for CV. Superior temporal cortex (h²_{CTh}=0.18, h²_{CSA}=0.30, h²_{CV}=0.26), precuneus (h²_{CTh}=0.16, h²_{CSA}=0.29, h²_{CV}=0.28) and pericalcarine (h²_{CTh}=0.15, h²_{CSA}=0.36, h²_{CV}=0.32) are among the most genetically determined regions.

The results of partitioned heritability analyses for global and regional CTh, CSA and CV with functional annotation and additionally with cell-type specific annotation are presented in Tables S27 and S28. For global CTh we found enrichment for super-enhancers, introns and histone marks. Repressors and histone marks were enriched for global CSA, and introns, super-enhancers and repressors for global CV. For regional CSA and CV the highest enrichment scores (>18) were observed for conserved regions.

Genetic correlation

We found high genetic correlation (r_g) between global CSA and global CV $(r_g=0.81, p=1.2\times10^{-186})$ and between global CTh and global CV $(r_g=0.46, p=1.4\times10^{-14})$, but not between global CTh and global CSA $(r_g=-0.02, p=0.82)$. Whereas genetic correlation

between CSA and CV was strong ($r_g > 0.7$) in most of the regions (Table S29 and Supplementary Figure 12), it was generally weak between CSA and CTh with $r_g < 0.3$, and ranged from 0.09 to 0.69 between CTh and CV. The postcentral and lingual cortices were the two regions with the highest genetic correlations between both CTh and CV, as well as CTh and CSA.

Genetic correlation across the various brain regions for CTh (Supplementary Figure 13, Table S30), CSA (Supplementary Figure 14, Table S30), and CV (Supplementary Figure 15, Table S32) showed a greater number of correlated regions for CTh and greater inter-regional variation for CSA and CV. Tables S33 - S35 and Supplementary Figures 16-18 show genome-wide genetic correlations between the cortical measures and anthropometric, neurological and psychiatric, and cerebral structural traits.

Discussion

In our genome-wide association study of up to 22,824 individuals for MRI determined cortical measures of global and regional thickness, surface area and volume, we identified 160 genome-wide significant associations across 19 chromosomes. Heritability was generally higher for cortical surface area and volume than for thickness, suggesting a greater susceptibility of cortical thickness to environmental influences. We observed strong genetic correlations between surface area and volume, but weak genetic correlation between surface area and thickness. We identified the largest number of novel genetic associations with cortical volumes, perhaps due to our larger sample size for this phenotype which was assessed in all 21 discovery samples.

It is beyond the scope of our study to discuss each of the 160 associations identified. However, broad patterns emerged showing that genes determining cortical structure are also often implicated in development of the cerebellum and brainstem (*KIAA0586, ZIC4, ENPP2*) as well as the neural tube (one carbon metabolism genes *DHFR* and *MSRBB3*, the latter also associated with hippocampal volumes³³). These genes determine development of not only neurons but also astroglia (*THBS1*) and microglia (*SALL1*). They determine susceptibility or resistance to a range of insults: inflammatory, vascular (*THBS1, ANXA1, ARRDC3-AS1*³⁴) and neurodegenerative (*C15orf53, ZIC4, ANXA1*), and have been associated with pediatric and adult psychiatric conditions (*THBS1*). At a molecular level, the wnt/β-catenin, TGF-β and sonic hedgehog pathways are strongly implicated. Gene-set-enrichment analyses revealed biological processes related to brain morphology and neuronal development.

There is a wealth of information in the supplementary tables that can be mined for a better understanding of brain development, connectivity, function and pathology. We highlight this potential by discussing in additional detail, the possible significance of 6 illustrative loci, 5 of

which, at 15q14, 14q23.1, 6q22.32, 17q21.31 and 3q24, associate with multiple brain regions at low p-values, while the locus at 8q24.12 identifies a plausible exonic variant. The Chr15q14 locus was associated with cortical thickness, surface area and volumes in the postcentral gyrus as well as with surface area or volume across 6 other regions in the frontal and parietal lobes. Lead SNPs at this locus were either intergenic between C15orf53 and C15orf54, or intergenic between C15orf54 and THBS1 (Thrombospondin-1). C15orf53 has been associated with an autosomal recessive form of spastic paraplegia showing intellectual disability and thinning of the corpus callosum (hereditary spastic paraparesis 11, or Nakamura Osame syndrome). Variants of THBS1 were reported to be related to autism³⁵ and schizophrenia³⁶. The protein product of *THBS1* is involved in astrocyte induced synaptogenesis³⁷, and regulates chain migration of interneuron precursors migrating in the postnatal radial migration stream to the olfactory bulb³⁸. Moreover, *THBS1* is an activator of TGFβ signaling, and an inhibitor of pro-angiogenic nitric oxide signaling which plays a role in several cancers and immune-inflammatory conditions. Variants at Chr14q23.1 were associated with cortical surface area and volume of all regions in the occipital lobe, as well as with thickness, surface area and volume of the middle temporal cortex, banks of the superior temporal sulcus, fusiform, supramarginal and precuneus regions, areas associated with discrimination and recognition of language or visual form. These variants are either intergenic between KIAA0586, the product of which is a conserved centrosomal protein essential for ciliogenesis, sonic hedgehog signaling and intracellular organization, and DACT1, the product of which is a target for SIRT1 and acts on the wnt/β-catenin pathway. KIAA0586 has been associated with Joubert syndrome, another condition associated with abnormal cerebellar development. Other variants are intergenic

between DACT1 and DAAM1 or intronic in DAAM1. DAAM1 has been associated with

occipital lobe volume in a previous GWAS.

Locus 6q22.32 contains various SNPs associated with cortical surface area and volume globally, and also within some frontal, temporal and occipital regions. The SNPs are intergenic between RSPO3 and CENPW. RSPO3 and CENPW have been previously associated with intracrania139,40 and occipital lobe volumes. RSPO3 is an activator of the canonical Wnt signaling pathway and a regulator of angiogenesis. Chr17q21.31 variants were associated with global cortical surface area and volume and with regions in temporal lobe. These variants are intronic in the genes *PLEKHM1*, *CRHR1*, *NSF* and WNT3. In previous GWAS analyses, these genes have been associated with general cognitive function⁴¹ and neuroticism^{42,43}. CRHR1, NSF and WNT3 were additionally associated with Parkinson's disease⁴⁴⁻⁴⁸ and intracranial volume^{39,40,49}. The NSF gene also plays a role in Neuronal Intranuclear Inclusion Disease⁵⁰ and CRHR1 is involved in anxiety and depressive disorders⁵¹. This chromosomal region also contains the MAPT gene, which plays a role in Alzheimer's disease, Parkinson's disease, and frontotemporal dementia^{52,53}. The protein product of the gene ZIC4 is a C2H2 zinc finger transcription factor that has an intraneuronal, non-synaptic expression and auto-antibodies to this protein have been associated with subacute sensory neuronopathy, limbic encephalitis and seizures in patients with breast, small cell lung or ovarian cancers. ZIC4 null mice have abnormal development of the visual pathway⁵⁴ and heterozygous deletion of the gene has also been associated with a congenital cerebellar (Dandy-Walker) malformation⁵⁵, thus implicating it widely in brain development as well as in neurodegeneration. C2H2ZF transcription factors are the most widely expressed transcription factors in eukaryotes and show associations with responses to abiotic (environmental) stressors. Another transcription factor, FOXC1, also associated with Dandy-Walker syndrome has been previously shown to be associated with risk of all types of ischemic stroke and with stroke severity. Thus, ZIC4 might be a biological target worth pursuing to ameliorate neurodegenerative disorders.

We found an exonic SNP within the gene *ENPP2* (Autotaxin) at 8q24.12 to be associated with insular cortical volume. This gene is differentially expressed in the frontal cortex of Alzheimer patients⁵⁶ and in mouse models of Alzheimer disease such as the senescence-accelerated mouse prone 8 strain (SAMP8) mouse. Autotaxin is a dual-function ectoenzyme, which is the primary source of the signaling lipid, lysophosphatidic acid. Besides Alzheimer disease, changes in autotaxin/lysophosphatidic acid signaling have also been shown in diverse brain related conditions such as intractable pain, pruritus, glioblastoma, multiple sclerosis and schizophrenia. In the SAMP8 mouse, improvements in cognition noted after administration of LW-AFC, a putative Alzheimer remedy derived from the traditional Chinese medicinal prescription 'Liuwei Dihuang' decoction, are correlated with restored expression of four genes in the hippocampus, one of which is *ENPP2*.

Among the other genetic regions identified, many have been linked to neurological and psychiatric disorders, cognitive functioning, cortical development and cerebral structure (detailed listing in Table S36).

Heritability estimates are, as expected, generally higher in the family-based Austrian Stroke Prevention-Family study (ASPS-Fam) than in the Rotterdam Study (RS) for CTh (average h²_{ASPS-Fam}=0.52; h²_{RS}=0.26), CSA (0.62 and 0.30) and CV (0.57 and 0.23). This discrepancy is explained by the different heritability estimation methods: pedigree-based heritability in ASPS-Fam versus heritability based on common SNPs that are in LD with causal variants⁵⁷ in RS.

Average heritability over regions is also higher for surface area and volume, than for thickness. The observed greater heritability of CSA compared to CTh is consistent with the previously articulated hypothesis, albeit based on much smaller numbers, that CSA is developmentally determined to a greater extent with smaller subsequent decline after young

adulthood, whereas CTh changes over the lifespan as aging, neurodegeneration and vascular injuries accrue^{1,3}. It is also interesting that brain regions more susceptible to early amyloid deposition (e.g. superior temporal cortex, precuneus) have a higher heritability. We found no or weak genetic correlation between CTh and CSA, globally and regionally, and no common lead SNPs, which indicates that these two morphological measures are genetically independent, a finding consistent with prior reports^{25,26}. In contrast, we found strong genetic correlation between CSA and CV and identified common lead SNPs for CSA and CV globally, and in 12 cortical regions. Similar findings have been reported in a previous publication²⁶. The genetic correlation between CTh and CV ranged between 0.09 and 0.77, implying a common genetic background in some regions (such as the primary sensory postcentral and lingual cortices), but not in others. For CTh, we observed genetic correlations between multiple regions within each of the lobes, whereas for CSA and CV we found genetic correlations mainly between different regions of the occipital lobe. Chen et al⁵⁸ have also reported strong genetic correlation for CSA within the occipital lobe. There were also a few genetic correlations observed for regions from different lobes, suggesting similarities in cortical development transcended traditional lobar boundaries.

A limitation of our study is the heterogeneity of the MR phenotypes between cohorts due to different scanners, field strengths, MR protocols and MRI analysis software. Therefore, association results were combined using a sample-size weighted meta-analysis which does not provide overall effect estimates. Moreover, our sample comprises of mainly European ancestry, limiting the generalizability to other ethnicities. Strengths of our study are the population-based design, the large age range of our sample (12 – 90 years), use of three cortical measures as phenotypes of cortical morphometry, and the replication of our CTh and CSA findings in a large and independent cohort. In conclusion, we identified patterns of

heritability and genetic associations with various global and regional cortical measures, as well as overlap of MRI cortical measures with genetic traits and diseases that provide new insights into cortical development, morphology and possible mechanisms of disease susceptibility.

Methods

Study Population

The sample of this study consist of up to 22,824 participants from 20 population-based cohort studies collaborating in the Cohorts of Heart and Aging Research in Genomic Epidemiology (CHARGE) consortium⁵⁹ and the UK Biobank (UKBB)⁶⁰. All the individuals were stroke-and dementia free, aged between 12 and 90 years, and of European ancestry, except for ARIC AA with African ancestry. Table S1 provides population characteristics of each cohort and Supplementary Section 1 provides a short description of each study. Each study secured approval from institutional review boards or equivalent organizations, and all participants provided written informed consent. Our results were replicated using summary GWAS findings of 22,635 individuals from the Enhancing Neuroimaging Genetics through Metanalysis (ENIGMA) consortium³¹.

Genotyping and Imputation

Genotyping was conducted using various commercially available genotyping arrays across the study cohorts. Prior to imputation, extensive quality control was performed in each cohort. Genotype data were imputed to the 1000 Genomes reference panel⁶¹ (mainly phase 1, version 3) using validated software. Details on genotyping, quality control and imputation can be found in Table S2.

Phenotype Definition

This study investigated CTh, CSA and CV globally in the whole cortex and in 34 cortical regions. Global and regional CTh was defined as the mean thickness of the left and the right hemisphere in millimeter (mm). Global CSA was defined as the total surface area of the left

and the right hemisphere in mm², while regional CSA was defined as the mean surface area of the left and the right hemisphere in mm². Global and regional CV was defined as the mean volume of the left and the right hemisphere in mm³. The 34 cortical regions are listed in Table S3. High resolution brain magnetic resonance imaging (MRI) data was obtained in each cohort using a range of MRI scanners, field strengths and protocols. CTh, CSA and CV were generated using the Freesurfer software package^{62,63} in all cohorts except for FHSucd, where an in-house segmentation method was used. MRI protocols of each cohort can be found in Table S4 and descriptive statistics of CTh, CSA and CV can be found in Tables S5, S6 and S7.

Genome-wide associations, meta-analysis, replication and annotation

Based on a predefined analysis plan, each study fitted linear regression models to determine the association between global and regional CTh, CSA and CV and allele dosages of single nucleotide polymorphisms (SNPs). Additive genetic effects were assumed and the models were adjusted for sex, age, age², and if needed for study site and for principal components to correct for population stratification. Cohorts including related individuals calculated linear mixed models to account for family structure. Details on association software and covariates for each cohort are shown in Table S2. Models investigating regional CTh, CSA and CV were additionally adjusted for global CTh, global CSA and global CV, respectively. Quality control of the summary statistics shared by each cohort was performed using EasyQC⁶⁴. Genetic variants with a minor allele frequency (MAF) < 0.05, low imputation quality (R²<0.4), and which were available in less than 10,000 individuals were removed from the analyses. Details on quality control are provided in Supplementary Section 2.

We then used METAL⁶⁵ to perform meta-analyses using the z-scores method, based on p-values, sample size and direction of effect, with genomic control correction. We performed 10,000 permutation tests based on cortical measurements from Rotterdam Study to estimate

the number of independent tests. Based on the permutation test results, the genome-wide significance threshold was set a priori at $1.09 \times 10-9$ (= 5×10^{-8} /46). We used the clumping function in PLINK⁶⁶ (linkage disequilibrium (LD) threshold: 0.2, distance: 300kb) to identify the most significant SNP in each LD block.

For replication of our genome-wide significant CTh and CSA associations, we used GWAS meta-analysis results from the ENIGMA consortium³¹ for all SNPs that were associated at a p-value $< 5 \times 10^{-8}$ and performed a pooled meta-analysis. The p-value threshold for replication was set to 3.1×10^{-4} (nominal significance threshold (0.05) divided by total number of lead SNPs (160)). CV was not available in the ENIGMA results. The NHGRI-EBI Catalog of published GWAS⁶⁷ was searched for previous SNP-trait associations at a p-value of 5×10^{-8} of lead SNPs.

Regional association plots were generated with LocusZoom⁶⁸, and the chromosomal ideogram with PHENOGRAM (http://visualization.ritchielab.org/phenograms/plot).

Annotation of genome-wide significant variants was performed using the ANNOVAR software package⁶⁹ and the FUMA web application⁷⁰. FUMA eQTL mapping uses information from three data repositories (GTEx, Blood eQTL browser, and BIOS QTL browser) and maps SNPs to genes based on a significant eQTL association. We used a false discovery rate threshold (FDR) of 0.05 divided by number of tests (46) to define significant eQTL associations. Gene-based analyses, to combine the effects of SNPs assigned to a gene, and gene set analyses, to find out if genes assigned to significant SNPs were involved in biological pathways, were performed using MAGMA⁷¹ as implemented in FUMA. The significance threshold was set to 5.87×10⁻⁸ for gene-based analyses (FDR threshold (0.05) divided by number of genes (18,522) and number of independent tests (46)) and to 1.02×10⁻⁷ for the gene-set analyses (FDR threshold (0.05) divided by the number of gene sets (10,651) and by the number of independent tests (46)). Additionally, FUMA was used to investigate a

significant chromatin interaction between a genomic region in a risk locus and promoter regions of genes (250 bp upstream and 500 bp downstream of a TSS). We used an FDR of 1×10^{-6} to define significant interactions.

We investigated cis (<1Mb) and trans (>1 MB or on a different chromosome) expression quantitative trait loci (eQTL) for genome-wide significant SNPs in 724 post-mortem brains from the Religious Order Study and the Rush Memory and Aging Project (ROSMAP)^{72,73} stored in the AMP-AD database. The samples were collected from the gray matter of the dorsolateral prefrontal cortex. The significance threshold was set to 0.001 (FDR threshold (0.05) divided by the number of independent tests (46)).

For additional pathway analyses of genes that were mapped to independent lead SNPs by FUMA, we searched the InnateDB database⁷⁴. The STRING database⁷⁵ was used for visualizing protein-protein interactions. Only those protein subnetworks with five or more nodes are shown.

Heritability

Additive genetic heritability (h²) of CTh, CSA and CV was estimated in two studies: the Austrian Stroke Prevention Family Study (ASPS-Fam; n=365) and the Rotterdam Study (RS, n=4472). In the population based family study ASPS-Fam, the ratio of the genotypic variance to the phenotypic variance was calculated using variance components models in SOLAR⁷⁶. In case of non-normalty, phenotype data were inverse-normal transformed. In RS, SNP-based heritability was computed with GCTA^{77,78}. These heritability analyses were adjusted for age and sex.

Heritability and partitioned heritability based on GWAS summary statistics was calculated from GWAS summary statistics using LD score regression (LDSC) implemented in the ldsc tool (https://github.com/bulik/ldsc). Partitioned heritability analysis splits genome-wide SNP

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heritability into 53 functional annotation classes (e.g. coding, 3' UTR, promoter, transcription factor binding sites, conserved regions etc.) and additionally to 10 cell-type specific classes (e.g. central nervous system, cardiovascular, liver, skeletal muscle etc.) as defined by Finucane et al.⁷⁹ to estimate their contributions to heritability. The significance threshold was set to 2.05x10⁻⁵ (0.05/number of functional annotation classes (53) / number of independent tests (46)) for heritability partitioned on functional annotation classes and 2.05<10⁻⁶ (0.05/number of functional annotation classes (53) / number of independent tests (46)) for heritability partitioned on annotation classes and cell types.

Genetic correlation

LDSR genetic correlation⁸⁰ between CTh, CSA and CV was estimated globally and within each cortical region. The significance threshold was set to 7.35×10-4 (nominal threshold (0.05) divided by number of regions (34) and by number of correlations (CSA and CV, CSA and CTh). Genetic correlation was also estimated between all 34 cortical regions for CTh, CSA and CV, with the significance threshold set to 8.91×10-5 (nominal threshold (0.05) divided by number of regions (34) times the number of regions -1 (33) divided by 2 (half of the matrix). Additionally, the amount of genetic correlation was quantified between CTh, CSA and CV and physical traits (height, BMI), neurological and psychiatric diseases (e.g. Alzheimer's disease, Parkinson's disease), cognitive traits and MRI volumes (p-value threshold (0.05/46/number of GWAS traits). As recommended by the ldst tool developers, only HapMap3 variants were included in these analyses, as these tend to be well-imputed across cohorts.

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CHARGE

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Austrian Stroke Prevention Family (ASPS) / Austrian Stroke Prevention Family Study (ASPS-Fam)

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Table 1. Genome-wide significant associations (p_{Discovery} < 1.09×10-9) of regional cortical thickness (lowest p-value of each cortical region at each genomic locus)

Lobe	Region	Locus	Position	Lead SNP	Nearest Gene	Annotation	N	p Discovery	PReplication	ppooled
temporal	superior temporal	16q24.2	87225139	rs4843227	LOC101928708	intergenic	21887	2.79E-12	2.45E-05	2.31E-15
		17q21.31	44861003	rs199504	WNT3	intronic	21887	1.30E-10	1.17E-04	5.85E-13
	middle temporal	14q23.1	59072144	rs10782438	KIAA0586	intergenic	21559	2.17E-13	2.76E-08	8.99E-21
	inferior temporal	2q35	217332057	rs284532	SMARCAL1	intronic	21885	1.03E-09	2.64E-01	3.04E-07
	banksts	14q23.1	59074878	rs160458	KIAA0586(intergenic	18342	9.39E-10	2.42E-09	6.45E-18
parietal	superior parietal	16q24.2	87225101	rs9937293	LOC101928708	intergenic	21886	2.68E-14	1.64E-13	2.27E-27
		1q41	215141570	rs10494988	KCNK2	intergenic	21886	2.60E-12	3.66E-08	2.63E-19
	postcentral	15q14	39633904	rs2033939	C15orf54	intergenic	21885	1.17E-73	5.18E-68	7.73E-136
occipital	lateral occipital	5q14.1	79933093	rs245100	DHFR	intronic	21886	2.68E-11	3.77E-06	1.16E-15
	cuneus	14q23.1	59624317	rs4901904	DAAM1	intergenic	21885	4.02E-14	3.17E-10	2.88E-23
	insula	16q12.1	51449978	rs7197215	SALL1	intergenic	21560	1.45E-13	2.00E-02	6.42E-12
		9q31.3	113679617	rs72748157	LPAR1	intronic	21560	1.46E-10	1.38E-04	5.16E-13

N: number of individuals in meta-analysis; $p_{Discovery}$: p-value of discovery GWAS meta-analysis in CHARGE; $p_{Replication}$: p-value of replication meta-analysis in ENIGMA; p_{pooled} : p-value of pooled discovery and replication meta-analysis; in bold: $p_{Replication} < 3.1 \times 10^{-4}$ (=0.05/Nl, Nl=160, total number of lead SNPs); banksts: banks of the superior temporal sulcus.

Table 2. Genome-wide significant as social from (phistograph 4 1.09×10-9) of global and regional cortical surface area (lowest p-value of each cortical region at each genomic locus)

Superior frontal 6q22.32 126792095 rs11759026 MIR588 intergenic 18617 5.21E-22 1.45	3.55E-22 1.0 3.56E-08 1.2 66E-03 2.0 65E-06 8.0 NA NA 04E-27 1.2 63E-95 72E-02 6.9 64E-07 1.9 67E-04 1.9 63E-09 4.9	
Superior frontal 6q22.33 127204623 rs9375477 RSPO3 intergenic 18617 4.86E-13 1.60	36E-08 1.2 36E-03 2.0 35E-06 8.0 NA NA 34E-27 1.2 32E-11 1.2 32E-14 9.9 32E-07 1.7 32E-02 6.9 34E-07 1.9 34E-07 1.9	.23E-19 .05E-10 .07E-16 NA NA .20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
Superior frontal 6q22.33 127204623 rs9375477 RSPO3 intergenic 18617 4.86E-13 1.60	96E-03 2.0 95E-06 8.0 NA NA 94E-27 1.2 12E-11 1.2 13E-14 9.9 12E-07 1.7 12E-02 6.9 14E-07 1.9 15E-04 1.9 15E-04 1.9 15E-04 1.9 15E-04 1.9	.05E-10 .07E-16 NA NA .20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
Superior frontal 5q14.3 92187932 rs17669337 NR2F1-AS1 intergenic 18272 1.40E-11 2.05	NA NA NA NA NA NA SE-95 SE-95 SE-95 SE-96 SE-07 SE-02 SE-07 SE-02 SE-07 SE-02 SE-07 SE-02 SE-07 SE-04 SE-07 SE-04 SE-07 SE-04 SE-07 SE-04 SE-09	.07E-16 NA NA .20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
trontal frontal caudal middle frontal 6q22.32 126876580 rs9388500 RSPO3 intergenic 17891 2.35E-11 pars opercularis 5q23.3 128734008 rs12187568 ADAMTS19 intergenic 16632 1.19E-16 pars triangularis 3q24 147106319 rs2279829 ZIC4 UTR3 18265 6.32E-20 1.94 7q21.3 96175094 rs10458281 LOC100506136 intergenic 18265 1.15E-17 2.42 8.45E-precentral 15q14 39634222 rs1080066 C15orf54 intergenic 18267 109 2.53 6q15 92002569 rs9345124 MAP3K7 intergenic 18267 5.50E-11 2.73 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.73	NA NA 04E-27 1.2 12E-11 1.2 13E-95 13E-14 9.9 12E-07 1.7 12E-02 6.9 14E-07 1.9 17E-04 1.9 13E-09 4.9	NA NA .20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
pars opercularis 5q23.3 128734008 rs12187568 ADAMTS19 intergenic 16632 1.19E-16 pars triangularis 3q24 147106319 rs2279829 ZIC4 UTR3 18265 6.32E-20 1.94 7q21.3 96175094 rs10458281 LOC100506136 intergenic 18265 1.15E-17 2.42 precentral 15q14 39634222 rs1080066 C15orf54 intergenic 18267 109 2.53 6q15 92002569 rs9345124 MAP3K7 intergenic 18267 5.50E-11 2.73 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.73 contact 2	NA 04E-27 1.2 03E-95 03E-14 9.9 02E-07 1.7 02E-02 6.9 04E-07 1.9 07E-04 1.9 03E-09 4.9	NA .20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
pars opercularis 5q23.3 128734008 rs12187568 ADAMTS19 intergenic 16632 1.19E-16 pars triangularis 3q24 147106319 rs2279829 ZIC4 UTR3 18265 6.32E-20 1.94	34E-27 1.2 32E-11 1.2 33E-95 32E-07 1.7 72E-02 6.9 34E-07 1.9 37E-04 1.9 33E-09 4.9	.20E-45 .20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10
frontal pars triangularis 3q24 147106319 rs2279829 ZIC4 UTR3 18265 6.32E-20 1.94 7q21.3 96175094 rs10458281 LOC100506136 intergenic 18265 1.15E-17 2.42 8.45E- precentral 15q14 39634222 rs1080066 C15orf54 intergenic 18267 109 2.53 6q15 92002569 rs9345124 MAP3K7 intergenic 18267 5.50E-11 2.73 2p16.3 48274592 rs386645843 FBXO11 intergenic 18269 9.51E-12 8.42 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.73	33E-95 33E-95 32E-07 1.7 72E-02 6.9 34E-07 1.9 37E-04 1.9 33E-09 4.9	.20E-26 1.00E- 200 .91E-24 .71E-16 .96E-10 .92E-16
7q21.3 96175094 rs10458281 LOC100506136 intergenic 18265 1.15E-17 2.42	33E-95 33E-14 9.9 32E-07 1.7 22E-02 6.9 34E-07 1.9 37E-04 1.9 33E-09 4.9	1.00E- 200 .91E-24 .71E-16 .96E-10 .92E-16
precentral 15q14 39634222 rs1080066 C15orf54 intergenic 18267 109 2.53 6q15 92002569 rs9345124 MAP3K7 intergenic 18267 5.50E-11 2.73 2p16.3 48274592 rs386645843 FBXO11 intergenic 18269 9.51E-12 8.42 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.72	33E-95 32E-07 1.7 32E-02 6.9 34E-07 1.9 37E-04 1.9 33E-09 4.9	200 .91E-24 .71E-16 .96E-10 .92E-16
6q15 92002569 rs9345124 MAP3K7 intergenic 18267 5.50E-11 2.73 2p16.3 48274592 rs386645843 FBXO11 intergenic 18269 9.51E-12 8.42 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.73	73E-14 9.9 12E-07 1.7 72E-02 6.9 14E-07 1.9 17E-04 1.9 13E-09 4.9	.91E-24 .71E-16 .96E-10 .92E-16
2p16.3 48274592 rs386645843 FBXO11 intergenic 18269 9.51E-12 8.42 superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.72	72E-07 1.7 72E-02 6.9 74E-07 1.9 77E-04 1.9 73E-09 4.9	.71E-16 .96E-10 .92E-16
superior temporal 4q26 119249835 rs55699931 PRSS12 intronic 18269 2.08E-11 2.72	72E-02 6.9 64E-07 1.9 07E-04 1.9 83E-09 4.9	.96E-10 .92E-16
	54E-07 1.9 77E-04 1.9 33E-09 4.9	.92E-16
2022 2 150022691 re12009404 LVDDCD :::htm:// 10260 5.045.44 2.54	77E-04 1.9 33E-09 4.9	
2q23.2 150022681 rs13008194 LYPD6B intronic 18269 5.94E-11 2.54	3E-09 4.9	1
temporal middle temporal 6q22.32 126964510 rs4273712 RSPO3 intergenic 18269 6.93E-10 1.07		.99E-12
banksts 14q23.1 59072226 rs186347 <i>KIAA0586</i> intergenic 18265 4.11E-10 1.83		.93E-18
fusiform 17q21.31 44822662 rs199535 <i>NSF</i> intronic 18269 1.01E-13 1.1 4	L 4E-06 8.1	.13E-18
transverse temporal 2q23.2 150012936 rs2046268 LYPD6B intronic 18264 9.09E-10 3.21	21E-10 1.7	.78E-18
15q14 39632013 rs71471500 <i>C15orf54</i> intergenic 18270 3.85E-24 5.5 5	5E-19 5.8	.88E-41
superior parietal 19p13.2 13109763 rs68175985 NFIX intronic 17324 8.84E-11 2.68	8E-17 2.9	.90E-26
20q13.2 52448936 rs6097618 <i>SUMO1P1</i> intergenic 18267 1.78E-16	NA	NA
inferior parietal 12q14.3 65797096 rs2336713 <i>MSRB3</i> intronic 18267 1.24E-12 2.99	9E-12 2.8	.85E-23
2p25.2 4563477 rs669952 <i>LINC01249</i> intergenic 18267 4.47E-10 1.37	7E-08 4.7	.73E-17
15q14 39633904 rs2033939 <i>C15orf54</i> intergenic 18272 9.07E-27 1.61	1E-28 1.5	.59E-53
	'9E-14 3.4	.46E-26
parietal 3q24 147106319 rs2279829 <i>ZIC4</i> UTR3 18272 7.38E-12 4.2 4	24E-16 2.2	.29E-26
15q14 39634222 rs1080066 <i>C15orf54</i> intergenic 18265 5.65E-47 2.4 4	4E-36 1.8	.87E-80
postcentral 3q24 147106319 rs2279829 <i>ZIC4</i> UTR3 18265 1.90E-21 1.69	9E-26 2.9	.92E-46
9q21.13 76144318 rs67286026 <i>ANXA1</i> intergenic 18265 3.58E-12 8.0 4	14E-06 7.8	.82E-16
14q23.1 59628609 rs74826997 <i>DAAM1</i> intergenic 18270 2.40E-24 4.41	1E-18 4.5	.59E-40
precuneus 6q23.3 138866268 rs9376354 <i>NHSL1</i> intronic 18270 7.80E-13 4.12	2 E-08 7.2	.28E-19
3q26 190666643 rs1159211 <i>SNAR-I</i> intergenic 18270 4.49E-10 2.0 4	1.5	.59E-13
lateral occipital 14q23.1 59627631 rs2164950 DAAM1 intergenic 18269 3.04E-26 2.92	2E-15 2.2	.25E-38
lingual 14q23.1 59628679 rs76341705 <i>DAAM1</i> intergenic 18270 1.57E-20 8.67	7E-13 9.9	.96E-31
14q23.1 59625997 rs73313052 <i>DAAM1</i> intergenic 18267 1.90E-32 3.19	9E-15 2.9	.96E-43
cuneus 13q31.1 80191873 rs9545155 <i>LINC01038</i> intergenic 18267 5.15E-10 2.98	98E-05 3.9	.91E-13
14q23.1 59628679 rs76341705 <i>DAAM1</i> intergenic 18267 4.67E-24 2.5 6	6E-19 3.3	.35E-41
occipital 5q12.1 60117723 rs6893642 <i>ELOVL7</i> intronic 18267 1.40E-13 1.68	8E-08 6.2	.29E-20
3g13 11 104724787 rs971550 4/C4M intergenic 18267 2 18E-10 1 31	31E-06 4.4	.49E-15
Indrical caring		.32E-17
	1	.49E-18
		.05E-17
		.04E-14
insula 10q25.3 118704077 rs1905544 <i>SHTN1</i> intronic 17599 4.06E-12 3.65		

N: number of individuals in meta-analysis; p_{Discovery}: p-value of discovery GWAS meta-analysis in

CHARGE, p_{Replication}: p-value of replication meta-analysis in ENIGMA; -p_{pooled}: p-value of pooled discovery and replication meta-analysis; in bold: $p_{Replication} < 3.1 \times 10^{-4}$ (=0.05/Nl, Nl=160, total number of lead SNPs); banksts: banks of the superior temporal sulcus.

Table 3. Genome-wide significant associations (policovery 4 1.09 10-9 of global and regional cortical volume (lowest p-value of each cortical region at each genomic locus)

Lobe	Region	Locus	Position	Lead SNP	Nearest gene	Annotation	N	Discovery
2000	педіон	6q22.32	126792095	rs11759026	MIR588	intergenic	22410	6.31E-19
	global		44790203	rs169201	NSF	intronic	22784	2.11E-13
		· ·	43549608	rs149366495		intronic	22099	8.18E-13
		12q14.3	66358347	rs1042725	HMGA2	3'UTR	22784	7.04E-11
		12q23.2	102921296	rs11111293	IGF1	intergenic	22784	5.45E-10
		6q22	109002042	rs4945816	FOXO3	3'UTR	22784	8.93E-10
	superior frontal	5q14.3	92186429	rs888814	NR2F1-AS1	intergenic	22692	3.29E-13
	rostral middle frontal	15q14	39636227	rs17694988	C15orf54	intergenic	22793	3.15E-11
	caudal middle frontal	2q12.1	105460333	rs745249	LINC01158	ncRNA_intronic		2.35E-11
		6q22.32	127068983	rs853974	RSPO3	intergenic	22351	4.82E-11
		5q23.3	128734008	rs12187568	ADAMTS19	intergenic	20753	4.27E-18
	pars opercularis	15q14	39639898	rs4924345	C15orf54	intergenic	22758	1.97E-14
		3q24	147106319	rs2279829	ZIC4	UTR3	22759	3.16E-23
frontal		7q21.3	96196906	rs67055449	LOC100506136	intergenic	22759	4.03E-19
	pars triangularis	15q14	39633904	rs2033939	C15orf54	intergenic	22759	8.49E-14
		7q21.3	96129071	rs62470042	C7orf76	intronic	22759	7.38E-13
		6q15	91942761	rs12660096	MAP3K7	intergenic	22759	4.74E-10
	lateral orbitofrontal	14q22.2	54769839	rs6572946	CDKN3	intergenic	22801	2.29E-10
		15q14	39634222	rs1080066	C15orf54	intergenic	22699	5.84E-125
	precentral	10q25.3	118648841	rs3781566	SHTN1	intronic	22699	4.68E-11
	superior temporal	3q26.32	177296448	rs13084960	LINC00578	ncRNA_intronic	22681	1.12E-11
	banksts	14q23.1	59072226	rs186347	KIAA0586	intergenic	22727	1.15E-15
		14q23.1	59833172	rs1547199	DAAM1	intronic	22605	4.58E-10
temporal	fusiform	1p33	47980916	rs6658111	FOXD2	intergenic	22605	7.78E-10
	transverse temporal	2q23.2	150012936	rs2046268	LYPD6B	intronic	22786	2.55E-12
	parahippocampal	2q33.1	199809716	rs966744	SATB2	intergenic	22747	2.23E-10
	superior parietal	15q14	39633904	rs2033939	C15orf54	intergenic	22723	4.28E-23
		16q24.2	87225139	rs4843227	LOC101928708	intergenic	22723	1.16E-13
		19p13.2	13109763	rs68175985	NFIX	intronic	21777	3.27E-11
		5q15	92866553	rs62369942	NR2F1-AS1	ncRNA_intronic		4.32E-10
	inferior parietal	20q13.2	52448936	rs6097618	SUMO1P1	intergenic	22701	2.09E-17
		12q14.3	65797096	rs2336713	MSRB3	intronic	22701	2.47E-13
		3q13.11	104724634	rs971551	ALCAM	intergenic	22701	2.34E-10
	supramarginal	15q14	39632013	rs71471500	THBS1	intergenic	22645	9.71E-28
		14q23.1	59627631	rs2164950	DAAM1	intergenic	22645	3.59E-20
parietal		3q24	147106319	rs2279829	ZIC4	UTR3	22645	5.36E-18
	postcentral	15q14	39633904	rs2033939	THBS1	intergenic	22662	4.34E-133
		3q24	147106319	rs2279829	ZIC4	UTR3	22662	2.54E-17
		9q21.13	76144318	rs67286026	ANXA1	intergenic	22662	5.03E-11
		2q36.3	226563259	rs16866701	NYAP2	intergenic	22545	5.69E-11
	precuneus	14q23.1	59628609	rs74826997	DAAM1	intergenic	22803	4.85E-20
		3q28	190663557	rs35055419	OSTN	intergenic	22428	2.02E-10
		2p22.2	37818236	rs2215605	CDC42EP3	intergenic	22803	3.43E-10
		3q13.11	104713881	rs12495603	ALCAM	intergenic	22803	9.71E-10
	lateral occipital	14q23.1	59627631	rs2164950	DAAM1	intergenic	22799	6.89E-16
	lateral occipital	14q23.1	59625997	rs73313052	DAAM1	intergenic	22805	1.06E-20
	lingual	6q22.32	127089401	rs2223739	RSPO3	intergenic	22805	1.75E-10
	cuneus	14q23.1	59625997	rs73313052	DAAM1	intergenic	22799	4.59E-43
		14q25.1 11p15.3	12072213	rs11022131	DKK3	intergenic	22799	5.96E-12
occipital		1 -				-		
		13q31.1 14q23.1	80192236 59628679	rs9545156 rs76341705	LINC01068 DAAM1	intergenic	22799 22824	4.09E-10 1.39E-29
						intergenic		1
	pericalcarine	13q31.1	80191873	rs9545155	LINC01068	intergenic	22824	2.25E-13
		11p14.1	30876113	rs273594	DCDC5	intergenic	22824	3.51E-13
		1p13.2	113208039	rs12046466	CAPZA1	intronic	22824	2.36E-12

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	not certified by peer revie	1p33 a	ıv aµi†æbye⊛ngın_o dera	F66658141ND	4 POMPonational lice	enir the proprint in	22824	3.85E-11
		11q22.3	104012656	rs1681464	PDGFD	intronic	22824	7.51E-11
		6q22.32	127096181	rs9401907	RSPO3	intergenic	22824	2.11E-10
		7p21.1	18904400	rs12700001	HDAC9	intronic	22824	2.12E-10
		5q12.1	60315823	rs10939879	NDUFAF2	intronic	22824	2.92E-10
	caudal anterior cingulate	5q14.3	82852578	rs309588	VCAN	intronic	22748	2.60E-10
	insula	11q23.1	110949402	rs321403	C11orf53	intergenic	22543	9.58E-12
		8q24.12	120596023	rs10283100	ENPP2	exonic	21481	8.29E-11

 $N: number \ of \ individuals \ in \ meta-analysis; \ p_{Discovery}: \ p\text{-value} \ of \ discovery} \ GWAS \ meta-analysis \ in$

CHARGE; banksts: banks of the superior temporal sulcus.

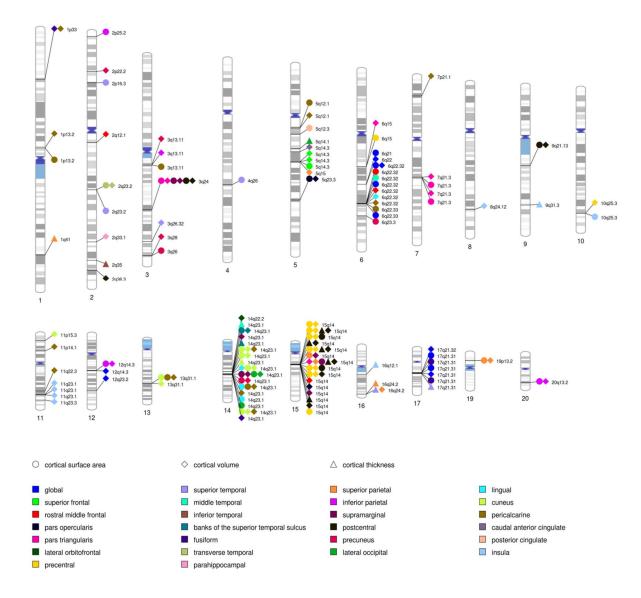


Figure 1. Chromosomal ideogram annotated with genome-wide significant associations $(p_{Discovery} < 1.09 \times 10^{-9})$ and corresponding genomic loci.

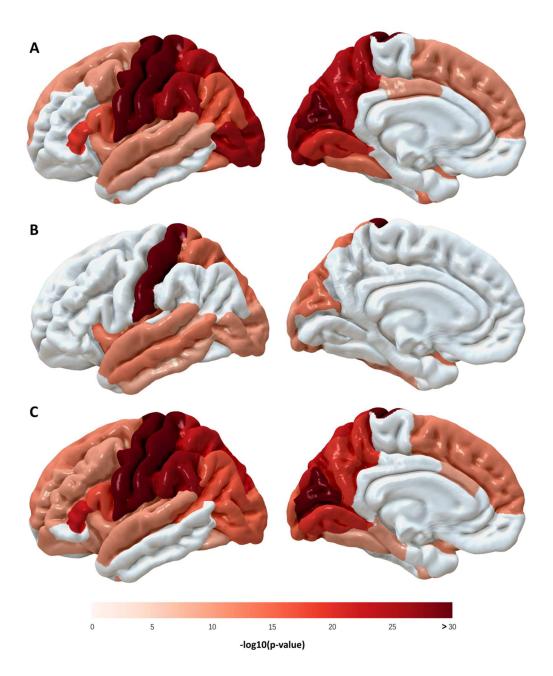


Figure 2. Lowest p-value of cortical surface area (A), thickness (B) and (C) volume of each cortical region.

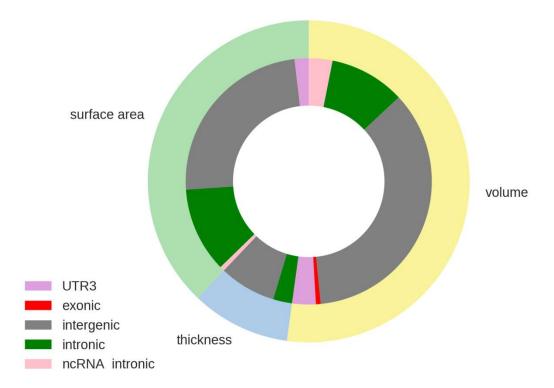


Figure 3. Proportion of functional annotation categories for global and regional cortical thickness, surface area and volume assigned by ANNOVAR.

Figure 4. Number of overlapping genes between FUMA eQTL mapping, FUMA chromatin interaction mapping, ANNOVAR chromosome positional mapping and MAGMA gene based analysis for all cortical regions combined for cortical surface area (A), thickness (B) and volume (C).