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Draft genome sequence of the *Saccharomyces cerevisiae Spy*Cas9 expressing strain IMX2600, a laboratory and platform strain from the CEN.PK lineage for cell-factory research

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ABSTRACT The biobased-economy aims to create a circular biotechnology ecosystem to transition from a fossil fuel-based to a sustainable industry based on biomass. For this, new microbial cell-factories are essential. We present the draft genome of the CEN.PK-derived *Saccharomyces cerevisiae Spy*Cas9 expressing strain (IMX2600), that serve as chassis of new cell-factories.

KEYWORDS *Saccharomyces cerevisiae*, CRISPR, biotechnology, cell factory, metabolic engineering

S trains belonging to the CEN.PK family were created as part of a multidisciplinary project funded by the Volkswagen Foundation in 1993–1994 (1). This lineage was chosen as a model for physiology and metabolic engineering research. With advent of CRISPR editing technology (2, 3) and increasing complexity of genetic engineering strategies that involve iteration of transformation rounds, chromosomal integration of *Spycas9* can accelerate strain engineering (4). To this end, the *Saccharomyces cerevisiae* strain IMX2600 (MATa *MAL2-8c SUC2 can1*Δ::*Spycas9*-natNT2) was developed from the prototrophic strain CEN.PK113-7D (MATa *MAL2-8c SUC2*) (5). IMX2600 have been used in constructing microbial cell factories (6–8). To ensure accurate sequencing analysis during construction and/or subsequent adaptive laboratory evolution strategy, a reference genome sequence of this platform strain is essential.

A -80°C stocked glycerol vial of the strain IMX2600 was inoculated and grown in 500 mL shake flasks with 100 mL YPD complex medium (10 g L⁻¹ Yeast extract, 20 g L^{-1} BactoPeptone, 20 g L^{-1} glucose). Total DNA was extracted using the YeaStar genomic DNA kit from Zymo Research (Zymo Research, Irvine, CA). Shotgun library preparation using Oxford_Nanopore Technologies' SQK-LSK109 kit (ONT, Oxford, UK) was sequenced on a MinION MK1B device with R10 flow cell. Raw FAST5 signal files were base-called using GPU Guppy 4.0.11 in high-accuracy mode. After filtering for quality and length, 974,343 reads were obtained, yielding 8.69 Gb (N50: 14,169 bp), representing ~724× coverage of a S. cerevisiae genome (Table 1). Canu version 2.0 with settings genomeSize = 12 m, useGrid = 0 and nanopore-raw (9) was used for *de novo* assembly yielding full-length chromosomes from telomere to telomere. Nucmer from the MUMmer package (Version 3.1) (10) was used to align and trim the 2µ plasmid and the mitochondrial DNA [nucmer (-maxmatch-nosimplify)]. For error correcting the assembled genome sequence, a 300 bp read length TruSeq PCR-Free Illumina library (Illumina, San Diego, CA) with a 550 bp insert-size yielding 2,793,193 paired reads for a total of 1.6 Gigabases (133-fold coverage) was sequenced on a MiSeq (Illumina). Reads quality was assessed with FastQC v0.11.5. The genome was polished by mapping the untrimmed Illumina reads with Burrows-Wheeler Aligner (BWA version 0.7.15-r1142-dirty; default parameters

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The authors declare no conflict of interest.

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Bioproject	PRJNA976676	https://www.ncbi.nlm.nih.gov/bioproject/PRJNA976676	
Biosample	SAMN35437089	https://www.ncbi.nlm.nih.gov/biosample/?term=SAMN35437089	
	Assembly size (Mb)	12.26	
	GC content (%)	38.26	
	N50 (bp)	914,509	
Experiment	Oxford nanopore	SRX20564515	
	Illumina	SRX20564514	
SRA	Oxford nanopore	https://trace.ncbi.nlm.nih.gov/Traces/?view=run_browser&acc=SRR24792116&display=metada	
	Illumina	https://trace.ncbi.nlm.nih.gov/Traces/?view=run_browser&acc=SRR24792117&display=metadata	
Assembly (annotated)	CP127195-CP127212	https://www.ncbi.nlm.nih.gov/nuccore?term=976676%5BBioProject%5D	

TABLE 1	IMX2600 assembly	y data and accession nur	nbers at NCBI re	epositories (h	ttps://www.ncbi.nlm.nih	.gov/
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used) (11, 12) to the assembly and further processed with SAMtools (version 1.3.1) (13, 14) and polished once by Pilon (version 1.18) with settings fix all (15). This yielded a genome assembly of 12.26 Megabase ($N_{50} = 915,509$ bp) composed of 16 chromosomes, a 86,554 bases mitochondrial genome and a 6,318 bases 2µ plasmid (Table 1) with a GC content of 38.26%. CHRXII that harbors the rDNA locus was assembled in one contig of 1,099,799 bp long and four smaller contigs (2x~8 kb +1x ~55 kb +1x ~100 kb) consisting exclusively of rDNA repeats that were not added to the assembly. The annotation of the polished assembly was performed using Funannotate v1.8.15 (https://github.com/nextgenusfs/funannotate) (16). The "predict" (Gene prediction) step was performed using the command (--species "Saccharomyces cerevisiae" --strain IMX2600— augustus_species saccharomyces_cerevisiae_S288C). Interproscan (version 5.25–64, with "-goterms") was applied in the "predict" step and used as input in the (Functional) "annotate" step. A total of 5,533 coding sequences and 287 tRNA's were identified.

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AUTHOR CONTRIBUTIONS

Marcel van den Broek, Data curation, Formal analysis, Methodology, Software | Raul A. Ortiz-Merino, Formal analysis, Methodology, Project administration, Software | Nicole X. Bennis, Investigation, Validation | Anna K. Wronska, Investigation, Validation | Else-Jasmijn Hassing, Investigation, Validation | Pascale Daran-Lapujade, Conceptualization, Project administration, Writing – original draft | Jean-Marc G. Daran, Conceptualization, Funding acquisition, Investigation, Methodology, Supervision, Writing – original draft

DATA AVAILABILITY

The genome sequencing and assembly of *Saccharomyces cerevisiae* strain IMX2600 have been deposited as Bioproject at NCBI under accession number PRJNA976676 (Table 1).

REFERENCES

- Entian KD, Kotter P. 2007. Yeast genetic strain and plasmid collections. Meth. Enzymol 36:629–666. https://doi.org/10.1016/S0580--9517(06)36025-4
- DiCarlo JE, Norville JE, Mali P, Rios X, Aach J, Church GM. 2013. Genome engineering in *Saccharomyces cerevisiae* using CRISPR-Cas systems. Nucleic Acids Res 41:4336–4343. https://doi.org/10.1093/nar/gkt135
- Jinek M, East A, Cheng A, Lin S, Ma E, Doudna J. 2013. RNA-programmed genome editing in human cells. eLife 2:e00471. https://doi.org/10.7554/ eLife.00471
- Mans R, van Rossum HM, Wijsman M, Backx A, Kuijpers NGA, van den Broek M, Daran-Lapujade P, Pronk JT, van Maris AJA, Daran JM. 2015. CRISPR/Cas9: a molecular swiss army knife for simultaneous introduction of multiple genetic modifications in *Saccharomyces cerevisiae*. FEMS Yeast Res 15:fov004. https://doi.org/10.1093/femsyr/fov004
- Wronska AK, van den Broek M, Perli T, de Hulster E, Pronk JT, Daran JM. 2021. Engineering oxygen-independent biotin biosynthesis in Saccharomyces cerevisiae. Metab Eng 67:88–103. https://doi.org/10. 1016/j.ymben.2021.05.006
- Bouwknegt J, Wiersma SJ, Ortiz-Merino RA, Doornenbal ESR, Buitenhuis P, Giera M, Müller C, Pronk JT. 2021. A squalene-hopene cyclase in *Schizosaccharomyces japonicus* represents a eukaryotic adaptation to sterol-limited anaerobic environments. Proc Natl Acad Sci U S A 118:e2105225118. https://doi.org/10.1073/pnas.2105225118
- Perli T, Vos AM, Bouwknegt J, Dekker WJC, Wiersma SJ, Mooiman C, Ortiz-Merino RA, Daran JM, Pronk JT. 2021. Identification of oxygenindependent pathways for pyridine nucleotide and coenzyme A synthesis in anaerobic fungi by expression of candidate genes in yeast. mBio 12:e0096721. https://doi.org/10.1128/mBio.00967-21
- Postma ED, Hassing EJ, Mangkusaputra V, Geelhoed J, de la Torre P, van den Broek M, Mooiman C, Pabst M, Daran JM, Daran-Lapujade P. 2022. Modular, synthetic chromosomes as new tools for large scale engineering of metabolism. Metab Eng 72:1–13. https://doi.org/10.1016/j.ymben. 2021.12.013

- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive K-MER weighting and repeat separation. Genome Res 27:722–736. https://doi. org/10.1101/gr.215087.116
- Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. Genome Biol 5:R12. https://doi.org/10.1186/gb-2004-5-2-r12
- Li H. 2013. Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM.arXiv. Genomics. https://doi.org/10.48550/arXiv. 1303.3997
- Li H, Durbin R. 2010. Fast and accurate long-read alignment with burrows-wheeler transform. Bioinformatics 26:589–595. https://doi.org/ 10.1093/bioinformatics/btp698
- Danecek P, Bonfield JK, Liddle J, Marshall J, Ohan V, Pollard MO, Whitwham A, Keane T, McCarthy SA, Davies RM, Li H. 2021. Twelve years of SAMtools and BCFtools. Gigascience 10:giab008. https://doi.org/10. 1093/gigascience/giab008
- Li H, Handsaker B, Wysoker A, Fennell T, Ruan J, Homer N, Marth G, Abecasis G, Durbin R, Genome Project Data Processing S. 2009. The sequence alignment/map format and SAMtools. Bioinformatics 25:2078–2079. https://doi.org/10.1093/bioinformatics/btp352
- Walker BJ, Abeel T, Shea T, Priest M, Abouelliel A, Sakthikumar S, Cuomo CA, Zeng Q, Wortman J, Young SK, Earl AM. 2014. Pilon: an integrated tool for comprehensive microbial variant detection and genome assembly improvement. PLoS One 9:e112963. https://doi.org/10.1371/ journal.pone.0112963
- Palmer JM, Stajich J. 2020. Funannotate V1.8.1: eukaryotic genome annotation. Zenodo 16. https://doi.org/10.5281/zenodo.1134477