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Article

Immobilization of *Arabidopsis thaliana* Hydroxynitrile Lyase (*AtHNL*) on EziG Opal

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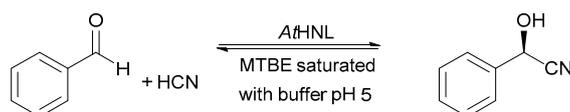


Abstract: *Arabidopsis thaliana* hydroxynitrile lyase (*AtHNL*) catalyzes the selective synthesis of (*R*)-cyanohydrins. This enzyme is unstable under acidic conditions, therefore its immobilization is necessary for the synthesis of enantiopure cyanohydrins. EziG Opal is a controlled porosity glass material for the immobilization of His-tagged enzymes. The immobilization of His₆-tagged *AtHNL* on EziG Opal was optimized for higher enzyme stability and tested for the synthesis of (*R*)-mandelonitrile in batch and continuous flow systems. *AtHNL*-EziG Opal achieved 95% of conversion after 30 min of reaction time in batch and it was recycled up to eight times with a final conversion of 80% and excellent enantioselectivity. The EziG Opal carrier catalyzed the racemic background reaction; however, the high enantioselectivity observed in the recycling study demonstrated that this was efficiently suppressed by using citrate/phosphate buffer saturated methyl-tert-butylether (MTBE) pH 5 as reaction medium. The continuous flow system achieved 96% of conversion and excellent enantioselectivity at 0.1 mL min⁻¹. Lower conversion and enantioselectivity were observed at higher flow rates. The specific rate of *AtHNL*-EziG Opal in flow was 0.26 mol h⁻¹ g_{enzyme}⁻¹ at 0.1 mL min⁻¹ and 96% of conversion whereas in batch, the immobilized enzyme displayed a specific rate of 0.51 mol h⁻¹ g_{enzyme}⁻¹ after 30 min of reaction time at a similar level of conversion. However, in terms of productivity the continuous flow system proved to be almost four times more productive than the batch approach, displaying a space-time-yield (STY) of 690 mol_{product} h⁻¹ L⁻¹ g_{enzyme}⁻¹ compared to 187 mol_{product} h⁻¹ L⁻¹ g_{enzyme}⁻¹ achieved with the batch system.

Keywords: *Arabidopsis thaliana*; hydroxynitrile lyase; oxynitrilase; His-tag; immobilization; batch; continuous flow

1. Introduction

Hydroxynitrile lyases (HNLs) are enzymes that catalyze the synthesis of enantiopure cyanohydrins (Scheme 1), known building blocks for the production of fine chemicals, pharmaceuticals and cosmetics [1–4]. HNL catalyzed reactions are faced with two problems, the chemical formation of racemic cyanohydrins and product racemization due to the reaction equilibrium [5]. These limitations can be overcome by performing the reactions in buffer saturated organic solvent and adjusting the pH to the lower limit accepted for HNLs [5,6]. These conditions are not the natural environment of HNLs, as they have to be stabilized for instance by immobilization on a suitable carrier.



Scheme 1. *AtHNL* catalyzed hydrocyanation of benzaldehyde yielding (*R*)-mandelonitrile.

Improved stability, activity and selectivity of immobilized enzymes have been reported earlier [7,8]. In addition, immobilization enables the increase of enzyme loading and facilitates recycling and downstream processing. To achieve the beneficial aspects mentioned before, the characteristics of enzyme and carrier must be considered. However, there is not a general method to immobilize an enzyme and its feasibility must be determined experimentally [9,10].

Immobilized metal ion chromatography (IMAC) is a widely used technique for the purification and immobilization of His-tagged enzymes. The enzyme immobilization is based on the affinity of divalent metal ions such as Zn^{2+} , Cu^{2+} , Ni^{2+} or Co^{2+} to the imidazole ring of histidines. Chelated Ni^{2+} on nitrilotriacetic acid (Ni-NTA) has turned out to be the most effective combination for the purification of His-tagged proteins [11]. However, nickel induced genotoxicity, carcinogenicity and immunotoxicity has been reported [12]. Hence, the development of a carrier with a non-toxic metal ion is highly desirable.

A new set of carriers (EziG, provided by EnginZyme AB, Stockholm, Sweden) containing non-toxic Fe^{3+} ($>10 \mu\text{mol g}^{-1}$) on its surface has been developed for the immobilization of His-tagged enzymes. These materials have a core made of controlled porosity glass (CPG) particles facilitating mass transfer from reactants and products to the material due to its interconnecting pore structure (circa 1.8 mL g^{-1}). In addition, its non-compressible non-swelling nature is an advantage compared to NTA agarose. The porous surface can be coated with an organic polymer to tailor carriers with different hydrophobic characteristics such as EziG Opal (hydrophilic), EziG Coral (hydrophobic) and EziG Amber (semi-hydrophobic). Given the hydrophilic surface of His₆-tagged *AtHNL* [13], its immobilization was performed on EziG Opal. Moreover, EziG Opal has been found to be suitable for reactions in organic solvents [14], a crucial property to enable the synthesis of enantiopure cyanohydrins together with low pH required in the reaction medium [5,15–19].

Some successful studies with different immobilized enzymes on EziG carriers have been reported earlier. An ω -transaminase was active in methyl-tert-butylether (MTBE) and a Baeyer–Villiger monooxygenase (BVMO) together with two cofactor-regenerating enzymes displayed increased stability [14]. An ω -transaminase from *Arthrobacter* sp. (AsR- ω TA) on EziG Amber was highly stable in batch and continuous flow systems [20]. On the other hand, when an old yellow enzyme (OYE) was immobilized on EziG Opal, poor recyclability was observed, and the initial conversion dropped to 56% after two reaction cycles [21]. The loss of activity of OYE was assumed to be due to enzyme leaching and/or deactivation of the enzyme. Likewise, the enzyme arylmalonate decarboxylase (AMDase) presented a significant loss of activity during recycling studies on all EziG carriers [22]. The loss of activity of AMDase activity on EziG carriers was explained to be due to enzyme leaching because of the lower strength of the coordinate bond and due to local pH changes by the acidic product of the reaction.

Enzyme catalyzed reactions in flow are gaining attention due to improved productivity, easier downstream processing and efficiency of scale-up compared to batch systems. Reduced reaction times and enhanced selectivity are reported benefits of performing reactions in flow [23–26]. On top of all these benefits mentioned before, continuous flow reactions allow to optimize resource utilization, reduce reaction volumes and consequently achieve waste reductions and lower energy consumption [27]. Furthermore, they allow handling of toxic and reactive reagents such as cyanide [28] in a safer manner.

The aim of this study is to immobilize *AtHNL* on EziG Opal based on the His-tag/ Fe^{3+} affinity and compare its performance for the synthesis of (*R*)-mandelonitrile with the earlier reported successful immobilization of *AtHNL* on Celite by adsorption [15]. Important parameters such as stability, specific rate and productivity were investigated in batch and flow systems.

2. Results and Discussion

AtHNL was recombinantly produced with a His₆-tag to enable its purification and immobilization by metal-ion affinity. It was successfully overexpressed in *E. coli* BL21(DE3) and purified displaying a specific activity of $136.5 \pm 3.2 \text{ U mg}^{-1}$. *AtHNL* was purified prior to its immobilization to avoid binding of other proteins with affinity to the EziG Opal carrier.

2.1. Batch Reactions

All batch reactions were performed with *AtHNL*-EziG Opal tightly packed into tea bags. Earlier research revealed that while the material of the bags had no influence on the conversion and enantioselectivity, it was essential to pack the bags tightly [16,18,29]. A magnetic stirrer was attached to the tea bag to enable the rotation of the immobilized enzyme and stirrer simultaneously. This set up avoids mechanical attrition of the carrier caused by the stirrer and facilitates the manipulation of the immobilized enzyme for recyclability studies without any loss of enzyme material. A leaching assay showed that *AtHNL* did not leach from EziG Opal carrier to the reaction medium (Figure S1). Similarly, no leaching had been reported previously for hydrocyanation reactions catalyzed by immobilized HNLs on siliceous carriers in general and *AtHNL* specifically [15,16,18,29].

Once it was established that EziG Opal is a suitable carrier for the immobilization of *AtHNL* (Figure S1), preliminary time studies using different enzyme loadings of *AtHNL*-EziG Opal for the synthesis of (*R*)-mandelonitrile were performed (Figure 1). The different enzyme loadings showed a huge difference in conversion and enantiopurity during four hours of reaction time. In these preliminary experiments, *AtHNL* was immobilized on EziG Opal by incubating an enzyme solution with the carrier in an orbital shaker (see Section 3.7 for details). The rotation enabled the enzyme to bind to the carrier but some precipitation was observed and it might explain the results in Figure 1. On the other hand, an earlier report [15] showed that *AtHNL* on Celite R-633 displayed near complete conversion and excellent enantioselectivity after 45 min using 5 mg mL^{-1} (circa 400 U) and MTBE saturated with citrate/phosphate buffer pH 5.5. The enzyme was immobilized by adsorption in that study. In addition, the successful immobilizations of *Prunus amygdalus* HNL (*PaHNL*) [16], *Manihot esculenta* HNL (*MeHNL*) [18] and *Granulicella tundricola* HNL (*GtHNL*) [29] on Celite were also performed by adsorbing all the enzyme solution into the carrier until saturation, which means that the enzyme solution is completely adsorbed into the carrier, just like in the case of Celite. All these results suggest that the immobilization of *AtHNL* on EziG Opal had to be optimized.

In order to optimize the immobilization of *AtHNL* on EziG Opal, the enzyme was immobilized by either incubating an enzyme solution in an orbital shaker or by adding it dropwise to EziG Opal carrier in such a way that the carrier absorbs the enzyme solution completely, as in the case of Celite [15,16,18,29]. Additionally, the effect of drying *AtHNL*-EziG Opal, which might influence the enzyme performance, was investigated. For this, *AtHNL*-EziG Opal was used either immediately after its immobilization (wet *AtHNL*-EziG Opal) or after 16 h of drying under vacuum in a desiccator over silica gel. Figure 2 shows the effect of drying *AtHNL*-EziG Opal and the immobilization method on the bioconversions. The immobilization of *AtHNL* in an orbital shaker with subsequent drying had a large negative impact on the conversion and enantioselectivity for the synthesis of (*R*)-mandelonitrile (Figure 2, dotted line and diamonds). The reaction catalyzed by wet *AtHNL*-EziG Opal (Figure 2, dashed line and triangles) proceeded faster and with improved enantioselectivity (92% of conversion and 92% of enantioselectivity) as compared to the dried *AtHNL*-EziG Opal (43% of conversion and 63% of enantioselectivity). As drying proved to have a negative impact on the enzyme activity and enantioselectivity, two reactions with wet *AtHNL*-EziG Opal immobilized by either incubation in an orbital shaker or adsorption were performed. A faster reaction was observed when the enzyme was immobilized by adsorption (Figure 2, continuous line and crosses), comparable to the results with Celite [15]. Similar conversion (circa 95% in both cases) was obtained after 4 h of reaction time, but enantioselectivity was slightly better for the enzyme immobilized by adsorption (96% *ee*) as compared to the enzyme immobilized by incubation (92% *ee*). Surprisingly, the effect of drying on the

reaction rate was negligible for *At*HNL immobilized by adsorption on EziG Opal (Figure 2, dashed line and dots). Conversions of circa 96% were obtained for both dried and non-dried *At*HNL-EziG Opal immobilized by adsorption within 30 min and little change was observed in the following 3.5 h. A possible explanation is that the enzyme is immediately stabilized right after its adsorption into the pores of the carrier, thus it is capable to endure the mechanical stress caused by the orbital shaker as well as the drying.

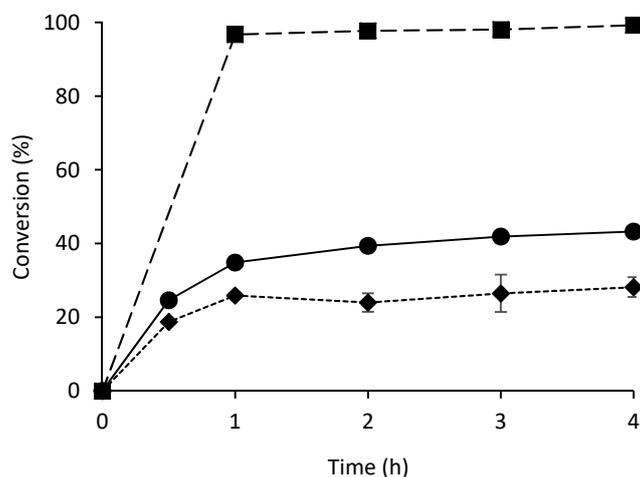


Figure 1. Synthesis of (*R*)-mandelonitrile using different enzyme loadings. Immobilization was performed by incubating enzyme and carrier in an orbital shaker and subsequent drying. Dashed line and squares (20 U mg^{-1} , final *ee* = 99%), solid line and dots (10 U mg^{-1} , final *ee* = 63%) and dotted line and diamonds (5 U mg^{-1} , final *ee* = 23%). Conditions: Ratio benzaldehyde:HCN in citrate/phosphate buffered MTBE, pH 5, 1:4, benzaldehyde (100 μL , 1 mmol), 2 mL HCN solution in citrate/phosphate buffered MTBE (1.5–2 M) pH 5, 27.5 μL (0.1 mmol) 1,3,5-triisopropylbenzene as internal standard (I.S.) and a teabag filled with *At*HNL immobilized on 60 mg EziG Opal. The reaction was stirred at 900 rpm at room temperature. Error bars correspond to the standard deviation of duplicates ($n = 2$) HPLC samples of the single experiment.

The lower conversions observed when the enzyme was immobilized by incubation might be explained by the loss of the *At*HNL dimeric structure caused by the rotation in an orbital shaker. In fact, some precipitation was observed after the incubation time. Earlier reports have shown that the enzyme stability is enhanced by higher oligomeric states [30]. Indeed, the ability of *Me*HNL to form tetramers in solution whereas *At*HNL forms dimers, explained the superior stability to higher temperatures and lower pH-values observed for *Me*HNL as compared to *At*HNL [31]. Similarly, *Me*HNL proved to be more stable than dimeric *Hevea brasiliensis* HNL (*Hb*HNL) for the synthesis of (*S*)-mandelonitrile in a monolith microreactor [17]. The formation of *Me*HNL tetrameric structures was attributed as the most likely reason for its higher stability.

Earlier studies [15] demonstrate a significant influence of the water content on enzyme activity of immobilized *At*HNL on Celite R-633, indicating that the minimal water content of *At*HNL-Celite for retaining enzymatic activity is 10% (*w/w*) of the immobilized enzyme. Additionally, the stability of *Me*HNL as CLEA or immobilized on Celite R-633 is highly dependent on the water entrapped in the carrier [18,32]. This effect can be ruled out here as buffer saturated MTBE was used as solvent.

Silica carriers, such as EziG Opal, are known to catalyze the chemical racemic background reaction [15–19,33,34] (Figures S2 and S3). However, the enantioselectivities achieved here in batch reactions under the optimized immobilization condition demonstrated the efficient suppression of this undesired reaction.

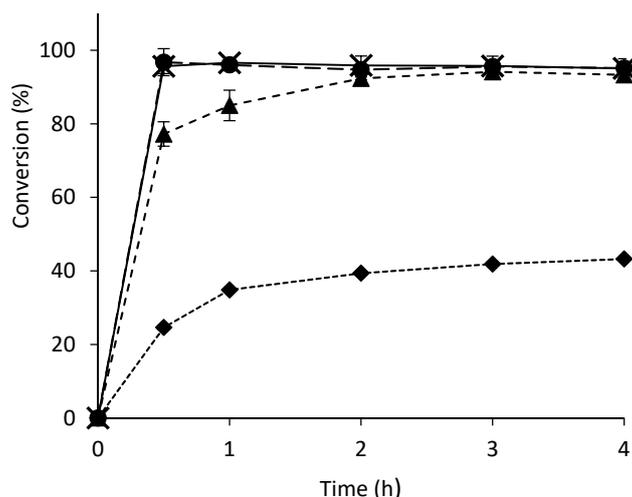


Figure 2. Effect of immobilization method and drying on the synthesis of (*R*)-mandelonitrile. Continuous line and crosses is the reaction with wet *AtHNL*-EziG Opal (adsorption), final *ee* = 96.2%; dashed line and dots is the reaction with dried *AtHNL*-EziG Opal (adsorption) final *ee* = 93.8%; dashed line and triangles is the reaction with wet *AtHNL*-EziG Opal (incubation in orbital shaker), final *ee* = 92.3% and dotted line and diamonds is the reaction with dried *AtHNL*-EziG Opal (incubation in orbital shaker), final *ee* = 63.3%. Conditions: Ratio benzaldehyde:HCN in citrate/phosphate buffered MTBE, pH 5, 1:4, benzaldehyde (100 μ L, 1 mmol), 2 mL HCN solution in citrate/phosphate buffered MTBE (1.5–2 M) pH 5, 27.5 μ L (0.1 mmol) 1,3,5-triisopropylbenzene as I.S. and a teabag filled with *AtHNL* immobilized on 60 mg EziG Opal. All reactions were performed with enzyme loading of 10 U mg^{-1} and the mol ratio of monomeric *AtHNL*: Fe^{3+} was 1:5. The reaction was stirred at 900 rpm at room temperature. Error bars of wet *AtHNL*-EziG Opal (adsorption), wet *AtHNL*-EziG Opal (incubation) and dried *AtHNL*-EziG Opal (incubation) correspond to the standard deviation of duplicate ($n = 2$) HPLC samples of the single experiment. Error bars of the reaction with dried *AtHNL*-EziG Opal (adsorption) correspond to the standard deviation of triplicate ($n = 3$) HPLC samples of the single experiment.

In addition to enzymatic activity and enantioselectivity, the stability of immobilized enzymes is crucial in biocatalytic applications. Indeed, one of the main objectives of enzyme immobilization is the potential for recycling the biocatalyst [4,6,35]. Since the best results for the synthesis of (*R*)-mandelonitrile were obtained with wet *AtHNL*-EziG Opal (10 U mg^{-1}) immobilized by adsorption, a recyclability study was performed under these conditions (Figure 3). In order to avoid enzyme overloading on the carrier which might lead to misinterpretations in the recyclability study, the mol ratio of monomeric *AtHNL*: Fe^{3+} used was 1:5, thus ensuring any enzyme deactivation is visible during the reaction cycles. Near complete conversion and excellent enantioselectivity (>99%) were observed during 7 cycles. When 10 U mg^{-1} of the enzyme were immobilized by incubation in an orbital shaker and subsequent drying, the recycling was unsuccessful (data not shown), whereas an enzyme loading of 20 U mg^{-1} led to a biocatalyst that could be recycled five cycles (Figure S4).

Figure 3 shows that EziG Opal enables to recycle *AtHNL* over several cycles with good conversion and enantioselectivity under the conditions of this study; accomplishing one of the main objectives of enzyme immobilization. Similarly, *AtHNL* on Celite displayed good conversion (>95%) and excellent enantioselectivity (>98% *ee*) during five consecutive reaction cycles [15]. Also, the successful recyclability of ω -transaminase from *Arthrobacter* sp. (*AsR*- ω TA) immobilized on EziG Amber (semi hydrophobic polymer surface) has been reported [20]. The immobilized *AsR*- ω TA (10 mg, 10% enzyme loading, $w w^{-1}$) was used for the kinetic resolution of *rac*- α -methylbenzylamine during 16 consecutive reaction cycles with excellent conversion and enantioselectivity. On the other hand, poor recyclability was recently reported [21] for the bioreduction of α -methyl-trans-cinnamaldehyde with a co-immobilized preparation of old yellow enzyme 3 (OYE3) and glucose dehydrogenase (GDH)

on EziG Opal (OYE3/GDH- EziG Opal). The conversion dropped to 56% after only two reaction cycles. However, it is worthy to mention that after 11 cycles (almost no conversion) the addition of GDH increased the conversion up to 30% suggesting that GDH was gradually deactivated or leached from the carrier over the reaction cycles. Also, the synthesis of enantiopure (*S*)-arylpropionate using arylmalonate decarboxylase (AMDase) immobilized on three EziG carriers with different surface hydrophobicity has been reported [22]. The best activity was obtained with the hydrophilic carrier (EziG Opal). Unfortunately, the enzyme was nearly fully deactivated after the second reaction cycle for the three EziG carriers. This significant loss of enzymatic activity was attributed to enzyme leaching or local pH shifts inside the porous carriers.

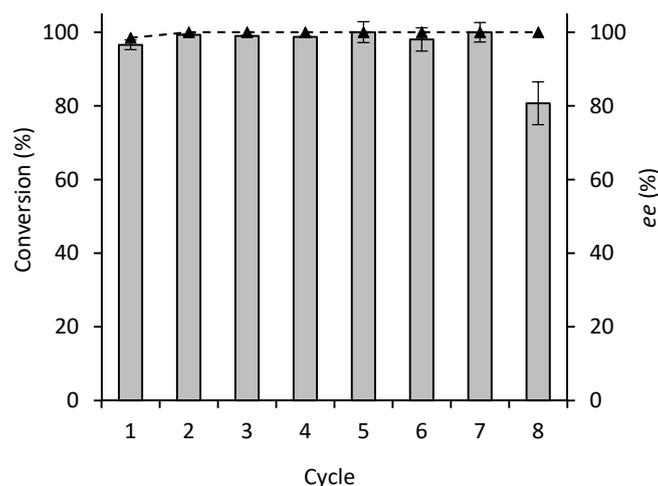


Figure 3. Recycling of wet *AtHNL*-EziG Opal (10 U mg^{-1}) in eight successive cycles. Immobilization was performed by adsorption. Conversion (bars), enantiomeric excess (dotted line and triangles). Conditions: Ratio benzaldehyde:HCN in citrate/phosphate buffered MTBE, pH 5, 1:4, benzaldehyde ($100 \mu\text{L}$, 1 mmol), 2 mL HCN solution in citrate/phosphate buffered MTBE ($1.5\text{--}2 \text{ M}$) pH 5, $27.5 \mu\text{L}$ (0.1 mmol) 1,3,5-triisopropylbenzene as I.S. and a teabag filled with *AtHNL* immobilized on 60 mg EziG Opal. Mol ratio of monomeric *AtHNL*: Fe^{3+} was 1:5. The reaction was stirred at 900 rpm at room temperature. The enzyme was washed for 1 min with 100 mM citrate/phosphate buffer saturated MTBE pH 5 after each cycle. Reaction time: 1 h . Error bars correspond to the standard deviation of duplicates ($n = 2$).

2.2. Continuous Flow Reactions

Immobilization enables the use of enzyme catalyzed synthesis reactions in continuous flow. Several advantages have been reported for this approach such as increased productivity, enhanced stability, reduced enzyme inhibition and easier downstream processing [26,36,37]. In addition, the reduced risk of manipulation of hazardous reagents, such as hydrogen cyanide, due to the smaller reaction volume used for the biocatalytic reactions is advantageous [28].

AtHNL was immobilized on EziG Opal in accordance with the optimized method (adsorption + wet *AtHNL*-EziG Opal) developed for batch reactions and tested in a continuous flow reactor (CFR). Figure 4 shows the synthesis of (*R*)-mandelonitrile at different flow rates. Near complete conversion and excellent enantioselectivity were achieved at 0.1 mL min^{-1} . An important decrease in enantioselectivity was observed at flow rates above 0.2 mL min^{-1} suggesting that *AtHNL* on EziG Opal suffers from the competing chemical background reaction catalyzed by the carrier. Indeed, the reduced enantioselectivity could be explained as the result of the carrier catalyzed chemical reaction (Figure S3). Water content and pH had a major impact on the synthesis of *rac*-mandelonitrile. Pure EziG Opal formed circa 5% of *rac*-mandelonitrile due to the chemical background reaction whereas the addition of $306 \mu\text{L}$ of phosphate buffer pH 5 (same volume used for the enzyme immobilization) increased its formation up to 26% at 0.1 mL min^{-1} (Figure S3).

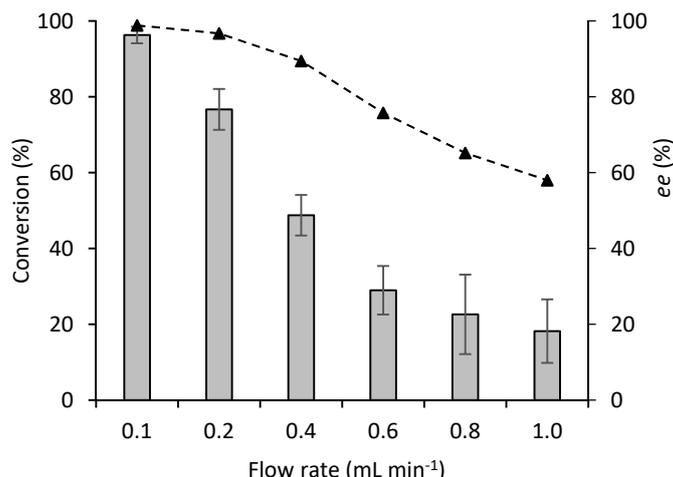


Figure 4. Synthesis of (*R*)-mandelonitrile using wet *AtHNL*-EziG Opal (10 U mg⁻¹) in a CFR. Immobilization was performed by adsorption. Conversion (bars), enantiomeric excess (dotted line and triangles). Conditions: benzaldehyde (0.5 M), HCN solution in citrate/phosphate buffered MTBE (1.5–2 M) pH 5, 1,3,5-triisopropylbenzene (50 mM, I.S.), a CFR with *AtHNL* immobilized on 150 mg EziG Opal. Mol ratio of monomeric *AtHNL*:Fe³⁺ was 1:4. Reactions were performed at room temperature. Error bars correspond to the standard deviation of triplicates ($n = 3$).

The stability of *AtHNL*-EziG Opal was evaluated in the synthesis of (*R*)-mandelonitrile at 0.1 mL min⁻¹ during 12 h on continuous operation. At this flow rate near complete conversion was achieved with a mol ratio of monomeric *AtHNL*:Fe³⁺ of 1:4, thus the robustness of the reaction system could be evaluated. Overall, *AtHNL*-EziG Opal displayed good conversion and high enantioselectivity during the stability study (Figure 5). Conversion of 74% and enantioselectivity of 89% respectively were achieved after 12 h of continuous operation. The decreased conversion and enantioselectivity after 12 h might be explained by enzyme deactivation due to the low pH 5 and the chemical reaction catalyzed by the carrier.

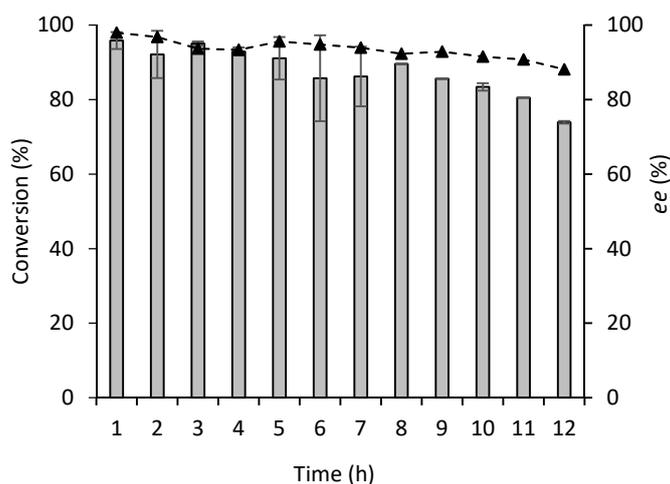


Figure 5. Stability of wet *AtHNL*-EziG Opal (10 U mg⁻¹) in continuous flow at 0.1 mL min⁻¹. Immobilization was performed by adsorption. Conversion (bars), enantiomeric excess (dotted line and triangles). Conditions: benzaldehyde (0.5 M), HCN solution in citrate/phosphate buffered MTBE (1.5–2 M) pH 5, 1,3,5-triisopropylbenzene (50 mM, I.S.), a CFR with *AtHNL* immobilized on 150 mg EziG Opal. Mol ratio of monomeric *AtHNL*:Fe³⁺ was 1:4. Reactions were performed at room temperature. Error bars correspond to the standard deviation of duplicates ($n = 2$) during the first 7 h. From hour 8, error bars correspond to the standard deviation of duplicates ($n = 2$) HPLC samples of the single experiment.

2.3. Comparison between Batch and Continuous Flow Systems

The comparison of the performance of the batch and continuous flow systems cannot be made based on conversions due to the different set ups used. To establish a clear comparison regarding the performance of *AtHNL*-EziG Opal in batch and continuous flow, the specific rate and space-time-yield (STY) at a similar level of conversion were calculated.

The specific rate at 0.1 mL min^{-1} (96% of conversion) was $0.26 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$, surprisingly, it is half of the specific rate calculated in batch. At 0.4 mL min^{-1} , a similar rate ($0.53 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$) was observed as compared to the batch system. However, the reduced conversion (49%) might make downstream processing more problematic. Higher flow rates did not further improve the specific rate. *AtHNL*-EziG Opal in batch displayed higher specific rate ($0.51 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$) compared to *AtHNL*-Celite [15] ($0.20 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$) at 96% of conversion. In both cases the reported conversion was achieved after 30 min reaction time.

Previously, *AtHNL* was immobilized on Celite R-633 [19] and tested for the synthesis of (*R*)-mandelonitrile, the reaction in flow using a packed bed reactor did not enhance the rate of the reaction as compared to the batch system; the best specific rate calculated for the continuous flow system was $0.04 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$ at 0.04 mL min^{-1} (85% of conversion), whereas the batch system showed $0.07 \text{ mol h}^{-1} \text{ g}_{\text{enzyme}}^{-1}$ (circa 91% of conversion). These results are circa six and seven-fold lower as compared to the best specific rate in flow (0.1 mL) and batch respectively reported in this study. In another study [17], the continuous flow synthesis of (*S*)-mandelonitrile with immobilized *HbHNL* on a siliceous monolith microreactor was 8 times faster as compared to the batch system and displayed a specific rate of $0.50 \text{ mmol min}^{-1} \text{ g}_{\text{enzyme}}^{-1}$ at circa 95% conversion and 0.2 mL min^{-1} . This result is twice the specific rate observed for *AtHNL*-EziG Opal at a similar level of conversion and might be explained by diffusion limitation due to the partial blockage of the pores of EziG Opal during the enzyme immobilization by adsorption. The irregular structure of the microchannels and mesopores found in monolith microreactors overcome this limitation.

The space-time-yield (STY) is a parameter commonly used to compare the productivity of reactors with different size. Batch systems often require rapid stirring to reduce mass transfer limitations that may shorten the lifetime of the immobilized enzyme. On the other hand, stirring is not required in flow, thus this problem is avoided, and better productivities can be achieved [17,25,29]. Indeed, the *AtHNL* on EziG Opal catalyzed synthesis of (*R*)-mandelonitrile in flow displayed a STY of $690 \text{ mol}_{\text{product}} \text{ h}^{-1} \text{ L}^{-1} \text{ g}_{\text{enzyme}}^{-1}$, whereas the batch approach led to only $187 \text{ mol}_{\text{product}} \text{ h}^{-1} \text{ L}^{-1} \text{ g}_{\text{enzyme}}^{-1}$ showing that the flow system greatly enhanced productivity. In batch, a similar productivity has been achieved previously with *AtHNL*-Celite ($150 \text{ mol}_{\text{product}} \text{ h}^{-1} \text{ L}^{-1} \text{ g}_{\text{enzyme}}^{-1}$) [15].

Comparing the results for *AtHNL* of this study with the literature reports again demonstrates the advantages of flow chemistry. The synthesis of (*S*)-mandelonitrile in a siliceous monolith microreactor using either *Hevea brasiliensis* HNL (11.3 mg total protein; 1120 U per monolith) or *Manihot esculenta* HNL (17.4 mg total protein; 1310 U per monolith) showed STYs of 555 and $405 \text{ mol}_{\text{product}} \text{ h}^{-1} \text{ L}^{-1} \text{ g}_{\text{enzyme}}^{-1}$ [17]. Recently, a ω -transaminase from *Arthrobacter* sp. (*AsR*- ω TA) was immobilized on EziG Amber (semi-hydrophobic carrier) for the kinetic resolution of *rac*- α -methylbenzylamine (*rac*- α -MBA) [20]. The enzyme was shown to be highly stable on this carrier and was able to perform the kinetic resolution of *rac*- α -MBA during 96 consecutive hours with excellent enantioselectivity (49% conversion and 99% *ee*). This flow system achieved a space time yield of $184 \text{ mol}_{\text{product}} \text{ h}^{-1} \text{ L}^{-1} \text{ g}_{\text{enzyme}}^{-1}$. The productivities reported in this study are comparable with the productivities reported for immobilized HNLs on siliceous carriers and other enzymes on EziG carriers.

3. Materials and Methods

3.1. Chemicals

Except when reported otherwise all chemicals were bought from Sigma Aldrich (Schnellendorf, Germany). Isopropanol and heptane were of HPLC grade ($\geq 99\%$) and used as HPLC solvents.

1,3,5-triisopropylbenzene (97%) was from Fluka Chemie (Buchs, Switzerland). Potassium cyanide (KCN, 97%) from J.T. Baker (Deventer, The Netherlands) was used as cyanide source in the HCN solution. (\pm)-Mandelonitrile from Acros Organics (Geel, Belgium) was purified by flash chromatography (PE/MTBE 9:1/3:7).

3.2. Heterologous Expression of *Arabidopsis Thaliana* HNL (*AtHNL*)

pET28a-*AtHNL* expression plasmid containing the *AtHNL* gene (GenBank accession number AAN13041, EC:4.1.2.10) codon optimized for *E. coli* and with a polyhistidine tag (His₆-tag) (see Table S1) was obtained from the group of Martina Pohl (Institute of Bio- and Geosciences, Jülich, Germany). *E. coli* BL21(DE3) was transformed with the expression plasmid for the production of the His-tagged *AtHNL*. A preculture was prepared by inoculating one single colony of *E. coli* BL21(DE3)-pET28a-*AtHNL* in 10 mL of Lysogeny Broth (LB) medium with kanamycin (40 $\mu\text{g mL}^{-1}$) and incubated overnight (Eppendorf/New Brunswick Scientific Incubator Shaker Excella E24 Series, Nijmegen, The Netherlands) at 37 °C, 180 rpm. Subsequently, this preculture was used for the inoculation of 1 L of Terrific Broth (TB) medium containing kanamycin (40 $\mu\text{g mL}^{-1}$) and incubated at 37 °C, 120 rpm. When the OD₆₀₀ reached 0.7–0.9 the protein expression was induced by adding 1 mL of 0.1 M isopropyl β -D-thiogalactoside (IPTG) per liter of culture (0.1 mM IPTG final concentration) and cultivation was continued at 25 °C, 180 rpm for 20 h.

Cells were harvested by centrifugation at 4 °C, 3600 \times g rpm during 20 min (Sorvall RC6, Thermo Fisher Scientific, Landsmeer, The Netherlands). The supernatant was discarded and the pellets were washed with 30 mL of 10 mM sodium phosphate buffer pH 7, frozen in liquid nitrogen and stored at –80 °C.

3.3. Enzyme Purification

The pellets containing *AtHNL* were resuspended in lysis buffer (10 mM potassium phosphate buffer pH 7.4 + DNase) and lysed in a cell disruptor (Constant Systems Ltd., Daventry, United Kingdom) at 1.5 kBar and 4 °C to avoid protein denaturation. The cell free extracts were collected by centrifugation at 48,000 \times g, 1 h, 4 °C (Sorvall RC6, Thermo Fisher Scientific, Landsmeer, The Netherlands). The enzyme was purified by using a NGC Chromatography system (Bio-Rad, Veenendaal, The Netherlands) by immobilized metal ion chromatography (IMAC) with chelating Ni²⁺ Sepharose (HiTrap Chelating HP 5 mL, GE Healthcare) according to the manufacturer [38]. 20 mM sodium phosphate + 0.5 M NaCl + 20 mM imidazole pH 7.4 was used for the enzyme binding and 20 mM sodium phosphate + 0.5 M NaCl + 0.5 M imidazole pH 7.4 was used for the enzyme elution.

The purified *AtHNL* was concentrated with a 10 kDa MWCO Amicon filter (Millipore, Amsterdam-Zuidoost, The Netherlands) and desalted with a PD-10 desalting column (Cytiva, Medemblik, The Netherlands) according to the supplier instructions [39].

3.4. Enzymatic Activity Assay

AtHNL activity was determined spectrophotometrically (Agilent Technologies Cary 60 UV-VIS, Amstelveen, The Netherlands) according to the literature [15] with slight modifications. The cleavage of *rac*-mandelonitrile into benzaldehyde and hydrogen cyanide was followed at 280 nm and 25 °C in 1 cm quartz glass cuvettes. Briefly, 1400 μL of 50 mM citrate/phosphate buffer pH 5 and 200 μL of enzyme solution (in 5 mM phosphate buffer pH 6.5) were mixed and incubated for 30 s at 25 °C. The reaction was started by adding 400 μL of 60 mM *rac*-mandelonitrile solution (80 μL of *rac*-mandelonitrile in 10 mL 3 mM citrate/phosphate buffer, pH 3.5). The enzymatic activity was calculated with the molar extinction coefficient of benzaldehyde ($\epsilon_{280} = 1.376 \text{ mM}^{-1} \text{ cm}^{-1}$) and the background reaction (performed without enzyme) was subtracted in the final calculation.

One unit of *AtHNL* activity is the amount of micromoles of *rac*-mandelonitrile converted per minute in citrate/phosphate buffer pH 5 at 25 °C.

3.5. Synthesis of Hydrogen Cyanide (HCN) Solution in MTBE

A HCN solution in MTBE was made according to earlier studies [15–19,29]. 25 mL MTBE and 10 mL MilliQ water were mixed in a 100 mL Erlenmeyer and kept at 0 °C. 0.1 mol potassium cyanide (6.51 g) was dissolved in the mixture and magnetically stirred for 15 min. 10 mL of 30% (v/v) HCl solution was added slowly and stirring was continued for 2 min. The HCN solution was allowed to reach room temperature (circa 20 °C). The organic and aqueous phases were separated using a separation funnel and the organic layer containing HCN was collected. The separation was performed twice more after adding 7 mL of MTBE each time. Finally, 5 mL of 50 mM citrate/phosphate buffer pH 5 was added to the organic fraction collected and it was stored in a dark bottle at 4 °C.

The HCN concentration was determined in accordance to the literature [40]. 1 mL of HCN solution was added to 5 mL of 2 M NaOH in a 25 mL Erlenmeyer. The mixture was stirred for 2 min. Potassium chromate was added as indicator. The solution was titrated with 0.1 M silver nitrate. The cyanide reacts 1:1 with the silver and precipitates.

3.6. Immobilization of *AtHNL* on EziG Opal by Adsorption

The immobilization of *AtHNL* on EziG Opal by adsorption was performed as described previously [16,18,29]. Given volumes of *AtHNL* solution were concentrated in Amicon filters with a 10 kDa MW cut-off, and subsequently added dropwise to 60 mg of EziG Opal (batch) or 150 mg of EziG Opal (flow). For batch reactions, *AtHNL*-EziG Opal was tightly packed in a regular paper tea bag [16,29] and either directly used for biocatalytic reactions or dried 16 h under vacuum in a desiccator over silica gel before packing. A magnetic stirrer was attached to the tea bags as reported earlier [16,29] to ensure tight packing and rapid stirring without mechanical attrition of the carrier. Reactions in flow were performed with wet *AtHNL*-EziG Opal (without drying and packing). The ratio of enzyme solution to carrier ($\mu\text{L}:\text{mg}$) was 2:1 in all cases to ensure that the enzyme solution was completely absorbed by the carrier. The immobilization of different enzyme units was achieved by determining the enzyme activity and adjusting the amount of enzyme solution before its concentration.

3.7. Immobilization of *AtHNL* on EziG Opal by Incubation

The immobilization of *AtHNL* on EziG Opal by incubation was performed according to the manufacturer (see the instruction manual in the supplementary information). 2 mL of enzyme solution with the required units to be immobilized was incubated with 60 mg of carrier in an orbital shaker (model RM:2M) at 30 rpm during 2 h at room temperature. The binding of the enzyme to the carrier was monitored by determining the protein concentration of the supernatant after immobilization using the BCA protein determination (Pierce BCA protein assay kit, Thermo Fisher Scientific, Landsmeer, The Netherlands) in accordance with the manufacturer instructions [41].

3.8. Synthesis of (*R*)-Mandelonitrile in Batch

Several (*R*)-mandelonitrile syntheses were performed with 60 mg of immobilized *AtHNL*-EziG Opal. The reaction conditions were as follows: 100 μL benzaldehyde (1 mmol), 27.5 μL 1,3,5-triisopropylbenzene (internal standard (I.S.)), 2 mL HCN in 50 mM citrate/phosphate buffered MTBE pH 5 (1.5–2 M), tea bag filled with 60 mg immobilized enzyme, 900 rpm and room temperature. The ratio benzaldehyde to HCN solution was 1:4. The mole ratio *AtHNL*: Fe^{3+} was 1:5.

3.9. Enzyme Recyclability in Batch

The enzyme recyclability was determined by several cycles of (*R*)-mandelonitrile synthesis according to [15,16,18,29]. The reaction conditions were as follows: benzaldehyde (100 μL , 1 mmol), 27.5 μL 1,3,5-triisopropylbenzene (I.S.), 2 mL HCN in 50 mM citrate/phosphate buffered MTBE pH 5 (1.5–2 M), tea bag filled with 60 mg immobilized enzyme, 900 rpm and room temperature. The ratio benzaldehyde to HCN solution was 1:4. The mol ratio *AtHNL*: Fe^{3+} was 1:5. Between each cycle the

immobilized enzyme was washed for 1 min with 50 mM citrate/phosphate buffered MTBE, pH 5, and stored at 4 °C in fresh citrate/phosphate buffered MTBE, pH 5.

3.10. Synthesis of (*R*)-Mandelonitrile in Continuous Flow

One milliliter stainless steel flow reactor (6.4 cm length, 0.45 cm inner diameter) was used for the continuous synthesis of (*R*)-mandelonitrile with 150 mg of immobilized *At*HNL on EziG Opal (10 U mg⁻¹). The reactor was filled with 100 mg of non-porous glass beads (1 mm diameter) and 150 mg of EziG Opal containing immobilized enzyme (final reaction volume = 0.3 mL). 10 cm of polytetrafluoroethylene (PTFE) tubing with 1.5 mm inner diameter was used to connect a syringe pump (Knauer, Germany) with the reactor. Initial conditions were as follows: 0.5 M benzaldehyde, 1.5–2 M HCN in 100 mM citrate/phosphate buffered MTBE, pH 5 and 50 mM 1,3,5-triisopropylbenzene as I.S.. The synthesis of (*R*)-mandelonitrile was evaluated at different flow rates (from 0.1 to 1 mL min⁻¹) by chiral HPLC. The mole ratio *At*HNL:Fe³⁺ was 1:4. The flow rate was checked at each sampling time by the difference of weight. Reactions were performed at room temperature. No significant pressure drop or increase was observed within the timeframe of the experiments.

3.11. Enzyme Stability in Continuous Flow

The stability of immobilized *At*HNL on EziG Opal (10 U mg⁻¹) was evaluated by performing a synthesis reaction during 12 h at 0.1 mL min⁻¹ on stream at room temperature. The mol ratio *At*HNL:Fe³⁺ was 25%. Samples were drawn at regular intervals and analyzed by chiral HPLC.

3.12. Analysis

Samples (10 µL) were taken at different times during the reaction run and added to 990 µL of heptane:2-propanol 95:5 in 1.5 mL Eppendorf tubes. A small amount of anhydrous magnesium sulphate (MgSO₄) was used to remove the water from the solution and the Eppendorf tubes were centrifuged at 13,000 × *g* rpm for 1 min. 850 µL of the supernatant was transferred to a 4 mL HPLC vial and 10 µL was injected into the HPLC (Chiralpak AD-H column, column size: 0.46 cm I.D × 25 cm). Heptane and 2-propanol were used as mobile phase with a flow rate of 1 mL min⁻¹ and the UV detector was set at 216 nm. The column temperature was set at 40 °C. The samples in the autosampler were maintained at 4 °C.

4. Conclusions

*At*HNL was successfully immobilized on EziG Opal by an optimized methodology. *At*HNL-EziG Opal was recycled up to seven times in batch with nearly complete conversion and excellent enantioselectivity. The continuous flow system displayed excellent conversion and enantioselectivity at 0.1 mL min⁻¹ and allowed to increase four times the productivity for the synthesis of (*R*)-mandelonitrile as compared to the batch system.

Supplementary Materials: The following information is available online at <http://www.mdpi.com/2073-4344/10/8/899/s1>, Figure S1: Leaching assay of *At*HNL-EziG Opal, Figure S2: Blank reaction in batch, Figure S3: Blank reaction in flow, Figure S4. Recycling of *At*HNL-EziG Opal (20 U mg⁻¹) in eight successive cycles. Figure S5. HPLC detection of benzaldehyde and 1,3,5-triisopropylbenzene during 8 h of incubation, Table S1: *At*HNL gene and Amino acid sequences.

Author Contributions: U.H. conceptualized and supervised the study; J.C., T.L. and M.A. performed the experiments; M.L. and P.B. developed the *At*HNL overexpression and purification method; P.-L.H., L.G. and U.H. reviewed the manuscript and supervised the study. J.C. wrote and edited the manuscript. All authors have read and agreed to the published version of the manuscript.

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References

1. Dadashipour, M.; Asano, Y. Hydroxynitrile lyases: Insights into biochemistry, discovery, and engineering. *ACS Catal.* **2011**, *1*, 1121–1149. [[CrossRef](#)]
2. Steiner, K.; Glieder, A.; Gruber-Khadjawi, M. Cyanohydrin formation/Henry reaction. In *Science of Synthesis: Biocatalysis in Organic Synthesis*; Georg Thieme Verlag: Stuttgart, Germany, 2015; Volume 2, pp. 1–30.
3. Lanfranchi, E.; Steiner, K.; Glieder, A.; Hajnal, I.; Sheldon, R.A.; van Pelt, S.; Winkler, M. Mini-Review: Recent developments in hydroxynitrile lyases for industrial biotechnology. *Recent Pat. Biotechnol.* **2013**, *7*, 197–206. [[CrossRef](#)]
4. Bracco, P.; Busch, H.; von Langermann, J.; Hanefeld, U. Enantioselective synthesis of cyanohydrins catalysed by hydroxynitrile lyases—A review. *Org. Biomol. Chem.* **2016**, *14*, 6375–6389. [[CrossRef](#)]
5. Faber, K. *Biotransformations in Organic Chemistry*, 7th ed.; Springer Nature: Basel, Switzerland, 2018; pp. 224–233. [[CrossRef](#)]
6. Hanefeld, U. Immobilisation of hydroxynitrile lyases. *Chem. Soc. Rev.* **2013**, *42*, 6308–6321. [[CrossRef](#)]
7. Zhu, Y.; Chen, Q.; Shao, L.; Jia, Y.; Zhang, X. Microfluidic immobilized enzyme reactors for continuous biocatalysis. *React. Chem. Eng.* **2020**, *5*, 9–32. [[CrossRef](#)]
8. Cantone, S.; Ferrario, V.; Corici, L.; Ebert, C.; Fattor, D.; Spizzo, P.; Gardossi, L. Efficient immobilisation of industrial biocatalysts: Criteria and constraints for the selection of organic polymeric carriers and immobilisation methods. *Chem. Soc. Rev.* **2013**, *42*, 6262–6276. [[CrossRef](#)] [[PubMed](#)]
9. Liese, A.; Hilterhaus, L. Evaluation of immobilized enzymes for industrial applications. *Chem. Soc. Rev.* **2013**, *42*, 6236–6249. [[CrossRef](#)] [[PubMed](#)]
10. Hanefeld, U.; Gardossi, L.; Magner, E. Understanding enzyme immobilisation. *Chem. Soc. Rev.* **2009**, *38*, 453–468. [[CrossRef](#)] [[PubMed](#)]
11. Block, H.; Maertens, B.; Spriestersbach, A.; Brinker, N.; Kubicek, J.; Fabis, R.; Labahn, J.; Schäfer, F. *Immobilized-Metal Affinity Chromatography (IMAC) in Methods in Enzymology*; Academic Press: Cambridge, MA, USA, 2009; pp. 439–473. [[CrossRef](#)]
12. Das, K.K.; Reddy, R.C.; Bagoji, I.B.; Das, S.; Bagali, S.; Mullur, L.; Khodnapur, J.P.; Biradar, M.S. Primary concept of nickel toxicity—An overview. *J. Basic Clin. Physiol. Pharmacol.* **2018**, *30*, 141–152. [[CrossRef](#)] [[PubMed](#)]
13. Andexer, J.N.; Staunig, N.; Eggert, T.; Kratky, C.; Pohl, M.; Gruber, K. Hydroxynitrile lyases with α/β -hydrolase fold: Two enzymes with almost identical 3D structures but opposite enantioselectivities and different reaction mechanisms. *ChemBioChem* **2012**, *13*, 1932–1939. [[CrossRef](#)] [[PubMed](#)]
14. Cassimjee, K.E.; Kadow, M.; Wikmark, Y.; Humble, M.S.; Rothstein, M.L.; Rothstein, D.M.; Bäckvall, J.-E. A general protein purification and immobilization method on controlled porosity glass: Biocatalytic applications. *Chem. Commun.* **2014**, *50*, 9134–9137. [[CrossRef](#)] [[PubMed](#)]
15. Okrob, D.; Paravidino, M.; Orru, R.V.A.; Wiechert, W.; Hanefeld, U.; Pohl, M. Hydroxynitrile lyase from *Arabidopsis thaliana*: Identification of reaction parameters for enantiopure cyanohydrin synthesis by pure and immobilized catalyst. *Adv. Synth. Catal.* **2011**, *353*, 2399–2408. [[CrossRef](#)]
16. Bracco, P.; Torrelo, G.; Noordam, S.; de Jong, G.; Hanefeld, U. Immobilization of *Prunus amygdalus* hydroxynitrile lyase on Celite. *Catalysts* **2018**, *8*, 287. [[CrossRef](#)]
17. van der Helm, M.P.; Bracco, P.; Busch, H.; Szymańska, K.; Jarzębski, A.; Hanefeld, U. Hydroxynitrile lyases covalently immobilized in continuous flow microreactors. *Catal. Sci. Technol.* **2019**, *9*, 1189–1200. [[CrossRef](#)]
18. Torrelo, G.; van Midden, N.; Stloukal, R.; Hanefeld, U. Immobilized hydroxynitrile lyase: A comparative study of recyclability. *ChemCatChem* **2014**, *6*, 1096–1102. [[CrossRef](#)]

19. Brahma, A.; Musio, B.; Ismayilova, U.; Nikbin, N.; Kamptmann, S.; Siegert, P.; Jeromin, G.E.; Ley, S.V.; Pohl, M. An orthogonal biocatalytic approach for the safe generation and use of HCN in a multistep continuous preparation of chiral *O*-Acetylcyanohydrins. *Synlett* **2015**, *27*, 262–266. [CrossRef]
20. Böhmer, W.; Knaus, T.; Volkov, A.; Slot, T.K.; Shiju, N.R.; Cassimjee, K.E.; Mutti, F.G. Highly efficient production of chiral amines in batch and continuous flow by immobilized ω -transaminases on controlled porosity glass metal-ion affinity carrier. *J. Biotechnol.* **2019**, *291*, 52–60. [CrossRef]
21. Tentori, F.; Bavaro, T.; Brenna, E.; Colombo, D.; Monti, D.; Semproli, R.; Ubiali, D. Immobilisation of old yellow enzymes via covalent or coordination bonds. *Catalysts* **2020**, *10*, 260. [CrossRef]
22. Aßmann, M.; Mügge, C.; Gaßmeyer, S.K.; Enoki, J.; Hilterhaus, L.; Kourist, R.; Liese, A.; Kara, S. Improvement of the process stability of Arylmalonate decarboxylase by immobilization for biocatalytic profen synthesis. *Front. Microbiol.* **2017**, *8*, 448. [CrossRef]
23. Yu, T.; Ding, Z.; Nie, W.; Jiao, J.; Zhang, H.; Zhang, Q.; Xue, C.; Duan, D.; Yamada, Y.M.A.; Li, P. Recent advances in continuous-flow enantioselective catalysis. *Chem. Eur. J.* **2020**, *26*, 5729–5747. [CrossRef]
24. Yoo, W.J.; Ishitani, H.; Saito, Y.; Laroche, B.; Kobayashi, S. Reworking organic synthesis for the modern age: Synthetic strategies based on continuous-flow addition and condensation reactions with heterogeneous catalysts. *J. Org. Chem.* **2020**, *85*, 5132–5145. [CrossRef] [PubMed]
25. Thompson, M.P.; Peñafiel, I.; Cosgrove, S.C.; Turner, N.J. Biocatalysis using immobilized enzymes in continuous flow for the synthesis of fine chemicals. *Org. Process Res. Dev.* **2019**, *23*, 9–18. [CrossRef]
26. Akwi, F.M.; Watts, P. Continuous flow chemistry: Where are we now? Recent applications, challenges and limitations. *Chem. Commun.* **2018**, *54*, 13894–13928. [CrossRef] [PubMed]
27. Sheldon, R.A.; Woodley, J.M. Role of biocatalysis in sustainable chemistry. *Chem. Rev.* **2018**, *118*, 801–838. [CrossRef]
28. Movsisyan, M.; Delbeke, E.I.P.; Berton, J.K.E.T.; Battilocchio, C.; Ley, S.V.; Stevens, C.V. Taming hazardous chemistry by continuous flow technology. *Chem. Soc. Rev.* **2016**, *45*, 4892–4928. [CrossRef]
29. Coloma, J.; Guiavarc'h, Y.; Hagedoorn, P.L.; Hanefeld, U. Probing batch and continuous flow reactions in organic solvents: *Granulicella tundricola* hydroxynitrile lyase (GtHNL). *Catal. Sci. Technol.* **2020**, *10*, 3613–3621. [CrossRef]
30. Vieille, C.; Zeikus, G.J. Hyperthermophilic enzymes: Sources, uses, and molecular mechanisms for thermostability. *Microbiol. Mol. Biol. Rev.* **2001**, *65*, 1–43. [CrossRef]
31. Guterl, J.K.; Andexer, J.N.; Sehl, T.; von Langermann, J.; Frindi-Wosch, I.; Rosenkranz, T.; Fitter, J.; Gruber, K.; Kragl, U.; Eggert, T.; et al. Uneven twins: Comparison of two enantiocomplementary hydroxynitrile lyases with α/β -hydrolase fold. *J. Biotechnol.* **2009**, 166–173. [CrossRef]
32. Paravidino, M.; Sorgedragger, M.J.; Orru, R.V.; Hanefeld, U. Activity and enantioselectivity of the hydroxynitrile lyase MeHNL in dry organic solvents. *Chem. Eur. J.* **2010**, *16*, 7596–7604. [CrossRef]
33. Effenberger, F.; Eichhorn, J.; Roos, J. Enzyme catalyzed addition of hydrocyanic acid to substituted pivalaldehydes—A novel synthesis of (*R*)-pantolactone. *Tetrahedron Asymmetry* **1995**, *6*, 271–282. [CrossRef]
34. Costes, D.; Wehtje, E.; Adlercreutz, P. Hydroxynitrile lyase-catalyzed synthesis of cyanohydrins in organic solvents: Parameters influencing activity and enantiospecificity. *Enzym. Microb. Technol.* **1999**, *25*, 384–391. [CrossRef]
35. Abdelraheem, E.M.M.; Busch, H.; Hanefeld, U.; Tonin, F. Biocatalysis explained: From pharmaceutical to bulk chemical production. *React. Chem. Eng.* **2019**, *4*, 1878–1894. [CrossRef]
36. Britton, J.; Majumdar, S.; Weiss, G.A. Continuous flow biocatalysis. *Chem. Soc. Rev.* **2018**, *47*, 5891–5918. [CrossRef] [PubMed]
37. Tamborini, L.; Fernandes, P.; Paradisi, F.; Molinari, F. Flow bioreactors as complementary tools for biocatalytic process intensification. *Trends Biotechnol.* **2018**, *36*, 73–88. [CrossRef]
38. Affinity Chromatography: Principles and Methods. Available online: www.sigmaaldrich.com/content/dam/sigma-aldrich/docs/Sigma-Aldrich/General_Information/1/ge-affinity-chromatography.pdf (accessed on 14 October 2019).
39. PD-10 Desalting Columns. Available online: <http://wwwuser.gwdg.de/~jjgrossh/protocols/protein-purification/PD10.pdf> (accessed on 14 October 2019).

40. Van Langen, L.M.; van Rantwijk, F.; Sheldon, R.A. Enzymatic hydrocyanation of a sterically hindered aldehyde. Optimization of a chemoenzymatic procedure for (*R*)-2-Chloromandelic Acid. *Org. Process. Res. Dev.* **2003**, *7*, 828–831. [CrossRef]
41. User Guide: Pierce BCA Protein Assay Kit. Available online: https://www.thermofisher.com/document-connect/document-connect.html?url=https%3A%2F%2Fassets.thermofisher.com%2F2FTFS-Assets%2F2FLSG%2Fmanuals%2FMAN0011430_Pierce_BCA_Protein_Asy_UG.pdf&title=VXNlciBHdWlkZTogUGllcmNlIEJDQSBQcm90ZWluIEFzc2F5IEtpdA (accessed on 15 October 2019).



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